Product sheet

OAW-42 Cells | 300304

General information

Description	The OAW-42 cell line was established from the ascites of a patient with ovarian cystadenocarcinoma. It has retained the ability to form free floating cysts in vitro, produces extracellular matrix, and shows a defined chemosensitivity pattern. It is a valuable cell line for studies on the biology of human ovarian cancer.
Organism	Human
Tissue	Ovary
Disease	Cystadenocarcinoma
Metastatic site	Ascites
Synonyms	OAW42, OAW 42

Characteristics

Age	46 years
Gender	Female
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	OAW-42 (Cytion catalog number 300304)
Biosafety level	1

Expression / Mutation

Ploidy status	Aneuploid
Karyotype	Hypotetraploid

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	25 to 30 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:4 to 1:6 is recommended
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	1 to 2 times per week
Freezing recovery	Fast. Allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	OAW-42 cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.
STR profile	**CSF1PO**: 11 **D13S317**: 11 **D16S539**: 12,13 **D5S818**: 11,12 **D7S820**: 8 **TH01**: 6,7 **TPOX**: 8,11 **vWA**: 15,16 **D3S1358**: 15,16 **D21S11**: 26 **D18S51**: 16,21 **Penta E**: 12 **Penta D**: 10 **D8S1179**: 13 **FGA**: 22,25
HLA alleles	**A***: 03:01:01, 30:02:01 **B***: 07:02:01, 18:01:01 **C***: 05:01:01, 07:02:01 **DRB1***: 01:01:01, 03:01:01 **DQA1***: 01:01:01, 05:01:01 **DQB1***: 02:01:01, 05:01:01 **DPB1***: 02:02:01G, 04:02:01G **E**: 01:03:02