

RT-112 Cells | 300324

General information

Description	This cell line was established by Dr. Carol Rigby, St. Paul's Hospital, London as described by Benham et al. in 1976 from a human bladder carcinoma.
Organism	Human
Tissue	Bladder
Disease	Carcinoma
Synonyms	RT 112, RT112

Characteristics

Age	Unspecified
Gender	Female
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Adherent

Regulatory Data

Citation	RT-112 (Cytion catalog number 300324)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1670

Biomolecular Data

Protein expression	p53 positive, Cytokeratine (4),5,(6), 7, 8, 13, 17, 18, 19, Desmoplakin
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Isoenzymes Yes, in nude mice

MSI-status Stable (MSS)

Handling

Culture Medium RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)

Subculturing Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Split ratio A ratio of 1:4 to 1:8 is recommended

Seeding density Start culture at 2 to 3 x 10⁴ cells/cm² and continue with a seeding density of 1 x 10⁴ cells/cm².

Fluid renewal 2 to 3 times per week

Freeze medium As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Quality Control & Molecular Analysis

Sterility Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,y
CSF1PO: 10,11
D13S317: 13,14
D16S539: 11,13
D5S818: 10,13
D7S820: 12,11
TH01: 7
TPOX: 8,11
vWA: 14,17
D3S1358: 15
D21S11: 27,3
D18S51: 15
Penta E: 12,16
Penta D: 10,11
D8S1179: 13,15
FGA: 23

HLA alleles

A*: '26:01:01
B*: '27:05:02
C*: '01:02:01
DRB1*: '01:01:01
DQA1*: '01:01:01
DQB1*: '05:01:01
DPB1*: '01:01:01
E: '01:01:01