Product sheet

BEAS-2B Cells | 300311

General information

Description	BEAS-2B is an immortalized cell line derived from the bronchial epithelium of a non-cancerous individual. This cell line was established by transforming human bronchial epithelial cells with an adenovirus 12-SV40 hybrid virus, which confers the cells with an extended lifespan while maintaining many of the morphological and functional characteristics typical of primary bronchial epithelial cells. BEAS-2B cells are widely used in respiratory disease research, particularly in studies related to the toxicological and pharmacological effects of inhalable substances, owing to their origin from the airway epithelium. The cell line exhibits a cobblestone morphology when cultured and retains certain critical features, such as the ability to metabolize xenobiotic compounds, making them highly relevant for studies on drug metabolism and respiratory toxicology. They have also been employed extensively in studies exploring cellular mechanisms of asthma, chronic obstructive pulmonary disease (COPD), and cancer. BEAS-2B cells respond predictably to cytokines, oxidative stress, and other stimuli typical of respiratory tract exposure to environmental agents. This makes them a valuable model for studying inflammation and oxidative stress mechanisms in pulmonary cells. As a tool in biomedical research, BEAS-2B cells are also frequently used to assess carcinogenic potential of airborne particles, where they serve as a model to understand the changes in airway epithelial cells following exposure to carcinogens. Their genetic makeup and susceptibility to genetic manipulation further enhance their utility in molecular biology experiments aimed at understanding gene expression and signaling pathways involved in lung diseases and cancer development.
Organism	Human
Tissue	Lung, Bronchus
Synonyms	Beas-2B, BEAS 2B, BEAS2B, Beas2B, Bronchial Epithelium transformed with Ad12-SV40 2B

Characteristics

Age	Age unspecified
Gender	Male
Morphology	Epithelial-like
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	BEAS-2B (Cytion catalog number 300311)

Biosafety level	1

Expression / Mutation

Viruses	Ad12-SV40 hybrid virus
Products	keratins, SV-40 T antigen

Handling

Culture Medium	Airway Epithelial Cell Basal Medium (PromoCell GmbH)
Medium supplements	Supplement the medium with Growth Medium Supplement Mix (PromoCell GmbH)
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.