Product sheet

SV-80 Cells | 300345

General information

Description	This SV40-transformed line was originally generated using cells which were derived from a skin biopsy of an adult female (strain A) by Todaro et al. in 1963, not from lung tissue of a five month old male fetus (strain C). After infection, the morphology of the growing colonies changed in that fibroblastic and epitheloid colony types were produced. The designation of SV-80 being of lung origin, and then retained, most probably was invalid. However, this cell line will characterized further in terms of p53 antigen and the presence of large T antigen.
Organism	Human
Tissue	Skin
Disease	Normal skin fibroblast (SV40-immortalized; non-tumorigenic)
Metastatic site	Not applicable (normal fibroblast line; not a tumor sample)
Applications	DNA repair research; SV40-immortalized fibroblast biology; cytogenetics; genotoxicity testing; normal human fibroblast reference for cancer comparator studies; SV40 large T antigen biology
Synonyms	SV-80, SV 80, SV-A clone 80, SV clone 80, Simian virus 80

Characteristics

Age	Adult
Gender	Female
Ethnicity	Caucasian
Morphology	Epithelial-like
Cell type	Fibroblast
Growth properties	Adherent

Regulatory Data

Citation	SV-80 (Cytion catalog number 300345)
Biosafety level	1

SV-80 Cells | 300345

NCBI_TaxID	9606
CellosaurusAccession	CVCL_0541
GMO Status	GMO-S1: This SV-80 human fibroblast line contains SV40 T-antigen sequences enabling immortalization for DNA repair and cytogenetics research. This classification applies only within Germany and may differ elsewhere.

Biomolecular Data

Tumorigenic	SMRV: Negative, as confirmed by Real-Time PCR
Karyotype	Modal number = 76, range = 52 to 87

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Doubling time	20 to 24 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1 to 5
Seeding density	3 to 5 × 10^3 cells/cm^2
Fluid renewal	1 to 2 times per week
Post-Thaw Recovery	Fast

Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.