Product sheet

ImWilms10T Cells | 300419

General information

Description	The imWilms10T cell line was immortalized from Wilms10 cells, a primary tumor cell line which was established in 2015 by the group of Dr. Brigitte Royer-Pokora from tissue of a 2-year-old female patient with triphasic stromal predominant Wilms tumor. Immortalization using the catalytic subunit of human telomerase 8hTERT) in conjunction with the mutant (U19dl89-97tsA58) SV40 large T antigen (LT) resulted in the cytogenetically stable cell line imWilms10.
Organism	Human
Tissue	Kidney
Disease	Wilms Tumor
Synonyms	ImWilms10 T, IM-WT-10

Characteristics

Age	2 years
Gender	Female
Ethnicity	Caucasian
Morphology	Spindle-shaped
Cell type	Wilms cells
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	ImWilms10T (Cytion catalog number 300419)
Biosafety level	1
Depositor	B. Royer-Pokora

Expression / Mutation

Mutational profile	WT1 mutation status: homozygous del WT1 within del11p13, LOH: no in 11p13 but UPD in 11p15, CTNNB1 mutation status: homozygous del TCT, p.DS45, UPD 3p

Handling

Culture Medium	MSCGM kit (from Lonza)
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	1 to 2 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile	**Amelogenin**: x,x **CSF1PO**: 11,12 **D13S317**: 12,12 **D16S539**: 9,10 **D5S818**: 10,12 **D7S820**: 11,12 **TH01**: 8,6 **TPOX**: 8,11 **vWA**: 15,18 **D3S1358**: 17,17 **D21S11**: 29,30 **D18S51**: 14,16 **Penta E**: 7,10 **Penta D**: 10,13 **D8S1179**: 10,15 **FGA**: 22,24
HLA alleles	**A***: 01:01:01, 11:01:01 **B***: 18:01:01, 27:05:02 **C***: 01:02:01, 12:03:01 **DRB1***: 01:01:01, 11:04:01 **DQA1***: 01:01:01, 05:05:01 **DQB1***: 03:01:01, 05:01:01 **DPB1***: 04:01:01G, 04:02:01G **E**: 01:01:01