### **Product sheet**





### **General information**

**Description**Animals were immunized with normal human cutaneous melanocytes. The antibody reacts with a heat-labile antigen on the same antigen. The antibody reacts with the M-24 antigen system.

**Organism** Mouse

**Tissue** Hematopoietic, hybridoma

**Synonyms** M138, M 138, M-24 (M138), M-24, L138

### **Characteristics**

Morphology Round cells

Cell type Lymphoblast

Growth properties

Suspension

## **Identifiers / Biosafety / Citation**

**Citation** L-138 (Cytion catalog number 400384)

Biosafety level 1

# **Expression / Mutation**

**Products**Monoclonal antibody (immunoglobulin, IgG1) against human cutaneous melanocytes (M-24 antigen system).
CLS does not warrant for antibody production of this cell line.

### **Handling**

**Culture** RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a) **Medium** 

Medium supplements

Supplement the medium with 10% FBS

**Subculturing** Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of  $2 \times 10^5$  cells/ml and keep the cell concentration within the range of  $1 \times 10^5$  to  $1 \times 10^6$  cells/ml for optimal growth.

#### **Product sheet**



## L-138 Cells | 400384

#### Fluid renewal

2 to 3 times per week

### Freeze medium

CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

### Handling of cryopreserved cultures

- 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
- 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
- 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
- 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
- 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
- 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
- 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
- 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

# Quality control / Genetic profile / HLA

#### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.