Product sheet

C643 Cells | 300298

General information

Description	The cell line C643 was established from a fine-needle biopsy of an anaplastic thyroid carcinoma of a 76-year-old man by Mark et al. in 1987. The patient died within 5 months after diagnosis. Demonstration of thyroglobulin mRNA ascertained a thyroid epithelial origin of the cell line. C643 cells emerge as a valuable tool for thyroid cancer research. These cells originated from human thyroid cancer tissue and represented metastatic PTC, FTC, and ATC. Their genetic makeup reflects the common mutations observed in thyroid cancer, such as alterations in BRAF, RAS, and PI3K genes, which activate critical signalling pathways. This makes C643 cells an ideal model for investigating the mechanisms involved in thyroid cancer development and progression. Furthermore, C643 cells are a crucial resource for testing potential targeted therapies. Their inclusion in preclinical studies can aid in identifying and evaluating novel compounds that specifically target the altered signalling pathways implicated in thyroid cancer. By accurately representing human thyroid cancer, C643 cells contribute to developing more effective treatments for patients with advanced thyroid cancer.
Organism	Human
Tissue	Thyroid gland anaplastic
Disease	Anaplastic thyroid carcinoma
Synonyms	C 643, C-643, c643

Characteristics

Age	76 years
Gender	Male
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Monolayer, adherent

Identifiers / Biosafety / Citation

Citation	C643 (Cytion catalog number 300298)

Biosafety level	1

Expression / Mutation

Tumorigenic	Yes, in nude mice

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:5 to 1:10 is recommended
Seeding density	1 x 10^4 cells/cm^2 will yield in a confluent layer in about 3 days
Fluid renewal	2 to 3 times per week
Freezing recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150?C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37?C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,y
CSF1PO: 10,11
D13S317: 8, 10
D16S539: 9, 13
D5S818: 11, 12
D7S820: 9, 12
TH01: 9.3, 10
TPOX: 11, 12
vWA: 15, 17
D3S1358: 15
D21S11: 28
D18S51: 14, 18
Penta E: 5, 15
Penta D: 9
D8S1179: 11, 13
FGA: 18, 21