

**Capan-1 Cells | 300143****General information**

<b>Description</b>	The cells express the cystic fibrosis transmembrane conductance regulator (CFTR) and secrete gastric type mucin.
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<b>Organism</b>	Human
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<b>Tissue</b>	Pancreas
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<b>Disease</b>	Ductal adenocarcinoma
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<b>Metastatic site</b>	Liver
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<b>Synonyms</b>	CaPan-1, CAPAN-1, Capan 1, CAPAN 1, Capan1, CAPAN1
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**Characteristics**

<b>Age</b>	40 years
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<b>Gender</b>	Male
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<b>Morphology</b>	Epithelial-like
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<b>Growth properties</b>	Adherent
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**Identifiers / Biosafety / Citation**

<b>Citation</b>	Capan-1 (Cyton catalog number 300143)
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<b>Biosafety level</b>	1
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**Expression / Mutation**

<b>Protein expression</b>	p53 negative
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<b>Antigen expression</b>	Blood Type A, Rh+
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<b>Isoenzymes</b>	Me-2, 1, PGM3, 1, PGM1, 1-2, ES-D, 1, AK-1, 1, G6PD, B, GLO-1, 1-2, Phenotype Frequency Product: 0.0311
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## Capan-1 Cells | 300143

**Tumorigenic** Form adenocarcinoma consistent with pancreatic duct carcinoma

**Products** Mucin

**Mutational profile** Capan-1 cells carry a homozygous Kras mutation in codon12: GGT(Gly) >GTT(Val)

**Karyotype** (P7) hypotriploid with abnormalities including dicentrics, breaks, acrocentric fragments, large submetacentric and subtelocentric chromosomes plus minute marker

## Handling

**Culture Medium** RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)

**Medium supplements** Supplement the medium with 10% FBS

**Passaging solution** Accutase

**Doubling time** 60 to 80 hours

**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

**Split ratio** A ratio of 1:2 to 1:4 is recommended

**Seeding density**  $2 \times 10^4$  cells/cm<sup>2</sup> will result in a 90% confluent monolayer in about 7 days

**Fluid renewal** Every 3 days

**Freezing recovery** After thawing, plate the cells at  $5 \times 10^4$  cells/cm<sup>2</sup> and allow the cells to recover from the freezing process and to adhere for at least 48 hours.

**Freeze medium** CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

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#### Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Quality control / Genetic profile / HLA

#### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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### STR profile

**Amelogenin:** x,x  
**CSF1PO:** 11  
**D13S317:** 9  
**D16S539:** 13,14  
**D5S818:** 11  
**D7S820:** 10,11  
**TH01:** 6  
**TPOX:** 8,11  
**vWA:** 16  
**D3S1358:** 15  
**D21S11:** 28,30  
**D18S51:** 12  
**Penta E:** 10,12  
**Penta D:** 9,13  
**D8S1179:** 14,16  
**FGA:** 24

### HLA alleles

**A\*:** 01:01:01, 30:01:01  
**B\*:** 13:02:01, 57:01:01  
**C\*:** 06:02:01  
**DRB1\*:** 07:01:01, 13:05:01  
**DQA1\*:** 02:01:01, 05:05:01  
**DQB1\*:** 02:02:01, 03:01:01  
**DPB1\*:** 03:01:01G, 04:01:01G  
**E:** 01:01:01