Product sheet

L Wnt-3A Cells | 305184

General information

Description	The L Wnt-3A cell line is a derivative of the L cells, originally derived from mouse fibroblast cells. This cell line is specifically engineered to stably express the Wnt-3A protein, a critical component of the Wnt signaling pathway. Wnt signaling is crucial for various developmental processes, including cell proliferation, differentiation, and migration. The stable expression of Wnt-3A in this cell line makes it a valuable tool for studying the molecular mechanisms underlying these biological processes, particularly in the context of cancer research, tissue regeneration, and embryonic development. Researchers often utilize the L Wnt-3A cell line to produce conditioned medium rich in Wnt-3A, which can then be used to activate Wnt signaling in other cell types. This application is especially beneficial in the study of stem cell biology and regenerative medicine, where Wnt signaling plays a pivotal role in maintaining stem cell pluripotency and promoting tissue repair. Additionally, the cell line serves as a model to investigate the dysregulation of Wnt signaling in various cancers, providing insights into potential therapeutic targets and treatments. Due to the robust and reliable expression of Wnt-3A, the L Wnt-3A cell line is widely used in laboratories to explore the effects of Wnt signaling on different cellular processes. It is an indispensable resource for scientists aiming to unravel the complexities of Wnt-mediated cellular functions and to develop novel strategies for modulating this pathway in disease contexts.
Organism	Mouse
Tissue	Subcutaneous connective tissue, areolar and adipose
Synonyms	L-Wnt-3A, L-Wnt3A, LWnt3A, LWnt-3A

Characteristics

Age	100 days
Gender	Male
Morphology	Fibroblast
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	L Wnt-3A (Cytion catalog number 305184)
Biosafety level	1

Expression / Mutation

Protein expression	Wnt-3A

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS, 0.4 mg/mL G-418
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1:2 to 1:4
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.