Product sheet

SCLC-22H growing culture | 330445

General information

Organism	Human
Tissue	Lung
Disease	Small cell carcinoma
Metastatic site	Pericardial effusion
Synonyms	SCLC22H

Characteristics

Age	46 years
Gender	Male
Ethnicity	Caucasian
Morphology	Floating cell aggregates, few single cells
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	SCLC-22H (Cytion catalog number 300445)
Biosafety level	1
Depositor	K?hler

Expression / Mutation

Tumorigenic	Yes, in nude mice
Reverse transcriptase	Negative

Karyotype	Modal number 43

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	The utilization of a passaging solution is not necessary when passaging cells that are cultured in suspension. The appropriate procedure is to dilute the cells in accordance with the indicated guidelines.
Subculturing	Dilute by transferring appropriate amounts of the suspension into fresh cell culture flasks. Start cultures at about 1 x 10^5 cells/ml.
Split ratio	A ratio of 1:2 to 1:6 is recommended
Seeding density	1 x 10^5 cells/ml
Fluid renewal	1 to 2 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	SCLC-22H cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.
Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in a 50 ml centrifuge tube. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Examine cell morphology and confluency using a microscope. Finally, incubate the flasks at 37 degrees Celsius for at least 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.
STR profile	**CSF1PO**: 10 **D13S317**: 12 **D16S539**: 12 **D5S818**: 11,12 **D7S820**: 11 **TH01**: 09. Mrz **TPOX**: 8,9 **vWA**: 17,18 **D3S1358**: 15 **D21S11**: 29,31.2 **D18S51**: 14,15 **Penta E**: 12,13 **Penta D**: 9 **D8S1179**: 12,13 **FGA**: 22
HLA alleles	**A***: 01:01:01, 32:01:01 **B***: 27:05:02, 51:01:01 **C***: 02:02:02 **DRB1***: 04:01:01, 09:01:02G **DQA1***: 03:01:01, 03:02:01 **DQB1***: 03:02:01, 03:03:02 **DPB1***: 02:01:02, 04:01:01 **E**: 01:01:01