

## Colo-205 Cells | 300380

### General information

**Description** COLO-205 cells express a 36,000 Dalton cell surface glycoprotein related to the GA733-2 tumor associated antigen.

**Organism** Human

**Tissue** Colon, Dukes' type D

**Disease** Colorectal adenocarcinoma

**Metastatic site** Ascites

**Synonyms** Colo 205, CoLo 205, COLO-205, COLO 205, COLO.205, Colo205, COLO205, Co 205, Colorado 205

### Characteristics

**Age** 70 years

**Gender** Male

**Morphology** Epithelial-like

**Growth properties** Adherent/suspension, loosely attached

### Identifiers / Biosafety / Citation

**Citation** Colo-205 (Cytion catalog number 300380)

**Biosafety level** 1

### Expression / Mutation

**Protein expression** CSAp- (Centriole and Spindle?Associated protein)

**Antigen expression** The cells are positive for keratin by immunoperoxidase staining.

**Isoenzymes** G6PD, B, PGM1, 1-2, PGM3, 1-2, 6PGD, A, ES-D, 1-2, PEP-D, 1

## Colo-205 Cells | 300380

**Tumorigenic** Yes, in nude mice

**Reverse transcriptase** Negative

**Products** Carcinoembryonic antigen (CEA) 1.5 to 4.1 ng/106 cells/10 days, keratin, interleukin 10 (IL-10, interleukin-10)

**Ploidy status** Aneuploid

### Handling

**Culture Medium** RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)

**Medium supplements** Supplement the medium with 10% FBS

**Doubling time** 20 to 25 hours

**Subculturing** Collect suspension cells in a 15 ml tube and carefully rinse the adherent cells using PBS without calcium and magnesium (3-5 ml PBS for T25, 5-10ml for T75 cell culture flasks). Add Accutase (1-2ml per T25, 2.5ml per T75 cell culture flask), the cell sheet must be covered completely. Incubate at ambient temperature for 10 minutes, then centrifuge the cells growing in suspension and the adherent cells together. Carefully resuspend the cells and dispense into new flasks which contain fresh medium.

**Split ratio** Subcultivation ratios of 1:2 to 1:10 are possible when all cells are pooled (suspended cells plus cells recovered after using Accutase)

**Seeding density**  $1 \times 10^4$  cells/cm<sup>2</sup>

**Fluid renewal** 2 to 3 times per week

**Freezing recovery** After thawing, plate the cells at  $5 \times 10^4$  cells/cm<sup>2</sup> and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

**Freeze medium** CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

### Colo-205 Cells | 300380

#### Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Quality control / Genetic profile / HLA

#### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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#### STR profile

**Amelogenin:** x,x  
**CSF1PO:** 11,12  
**D13S317:** 10,12  
**D16S539:** 12,13  
**D5S818:** 10,13  
**D7S820:** 9,10  
**TH01:** 8,9  
**TPOX:** 11  
**vWA:** 15  
**D3S1358:** 16  
**D21S11:** 30.2,33.2  
**D18S51:** 18  
**Penta E:** 13,15  
**Penta D:** 9,11  
**D8S1179:** 9,14  
**FGA:** 21,23

#### HLA alleles

**A\*:** 01:01:01, 02:01:01  
**B\*:** 07:02:01, 08:01:01  
**C\*:** 07:01:01, 07:02:01  
**DRB1\*:** 04:01:01, 13:01:01  
**DQA1\*:** 01:03:01  
**DQB1\*:** 06:03:01  
**DPB1\*:** 04:01:01  
**E:** 01:01:01, 01:03