Product sheet

DSL-6A-C1 Cells | 500166

General information

Description	DSL-6A/C1 is a pancreatic ductal cell line derived from the DSL-6 transplantable acinar cell carcinoma. The DSL-6 tumor was established in 1986 from a primary acinar cell carcinoma of the pancreas which developed in a male Lewis rat(DSL-101-79) that was given azaserine intraperitoneally. The cultured DSL-6A/C1 tumor cells initially produced amylase, but production of exocrine enzymes ceased after 1 to 2 weeks in culture. The cell line also lost structural and immunohistochemical acinar cell markers while acquiring duct cell markers during culture and regrafting. The DSL-6A/C1 cell line expresses the ductal marker cystic fibrosis transmembrane regulator (CFTR).
Organism	Rat
Tissue	Pancreas
Disease	Carcinoma, azaserine induced
Metastatic site	Ductal
Synonyms	DSL-6A/C1, DSL6A/C1

Characteristics

Age	2 years
Gender	Male
Morphology	Epithelial-like
Cell type	Acinar cells
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	DSL-6A-C1 (Cytion catalog number 500166)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes, in Lewis rats the cells produce solid tumors composed of ductlike structures surrounded by dense fibrous tissue

Handling

Culture Medium	Waymouth medium (We do not supply this product; please consider other suppliers. Please let us know if you need further assistance)
Medium supplements	Supplement the medium with 10% FBS, 2.0 mM L-glutamine
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:3 to 1:4 is recommended
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	2 times per week
Freezing recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150?C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37?C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Rat_D1Wox31: 104
Rat_D2Wox37: 156
Rat_D19Wox11: 232
Rat_D10Wox8: 266
Rat_D4Wox7: 157
Rat_D2Wox27: 207
Rat_D5Rat33: 122
Rat_D10Wox11: 171
Rat_D1Wox23: 210
Rat_D12Wox1: 406
Rat_D6Wox2: 104
Rat_D8Wox7: 182
Rat_D6Cebr1: 239
SRY: x,Y