

JAR Cells | 300221

General information

Description

The JAR cell line is a human choriocarcinoma cell line derived from trophoblastic cells of placental origin. This cell line is widely utilized in cancer research, particularly in studies related to gestational trophoblastic diseases and placental development. JAR cells exhibit characteristics typical of choriocarcinoma, including high levels of human chorionic gonadotropin (hCG) production, which makes them a valuable model for studying hormone regulation, placental biology, and the mechanisms underlying trophoblastic tumorigenesis.

JAR cells are known for their invasive properties and ability to proliferate rapidly, which mirrors the aggressive nature of choriocarcinomas in vivo. These cells are also used to investigate the interaction between trophoblastic cells and the maternal immune system, providing insights into immune evasion mechanisms. Additionally, JAR cells have been employed in studies of drug resistance and chemosensitivity, aiding in the development of therapeutic strategies against trophoblastic cancers. As a cell line derived from human tumors, JAR cells are strictly for in vitro research and are not suitable for any in vivo or therapeutic applications.

Organism

Human

Tissue

Placenta

Disease

Choriocarcinoma

Synonyms

Jar, JAr, JaR

Characteristics

Age

24 years

Gender

Female

Ethnicity

Caucasian

Morphology

Epithelial-like

Growth properties

Adherent

Regulatory Data

Citation

JAR (Cytion catalog number 300221)

Biosafety level

1

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NCBI_TaxID 9606**CellosaurusAccession** CVCL_0360**Biomolecular Data****Isoenzymes** G6PD, B, PGM1, 1-2, PGM3, 1-2, ES-D, 2, AK-1, 1, GLO-1, 1, Phenotype Frequency Product: 0.0002**Products** Estrogen, progesterone, hCG, human chorionic somatomammotropin (placental lactogen), hCG production averages 22.5 ng/ml after reculturing**Handling****Culture Medium** DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO₃ (Cytion article number 820400a)**Supplements** Supplement the medium with 10% FBS**Dissociation Reagent** Accutase**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Seeding density** 1×10^4 cells/cm²**Fluid renewal** Every 3 days**Post-Thaw Recovery** After thawing, plate the cells at 5×10^4 cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 24 hours.**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.