Product sheet

NFS-60 Cells | 400301

General information

Description	NFS-60 is a murine myeloblastic cell line established from leukemic cells obtained after infection of (NFS x DBA/2) F1 adult mice with Cas Br-M murine leukemia virus. NFS-60 cells are dependent on IL3 for growth and maintenance of viability in vitro. These cells are used to assay murine and human G-CSF. This bipotential murine hematopoietic cell line is responsive to IL-3, GM-CSF, G-CSF, and erythropoietin.
Organism	Mouse
Tissue	Blood
Disease	Leukemia
Synonyms	M-NFS-60, NFS 60, NFS60

Characteristics

Cell type	Lymphoblast
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	NFS-60 (Cytion catalog number 400301)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS, 33 IU/ml IL-3
Passaging solution	The utilization of a passaging solution is not necessary when passaging cells that are cultured in suspension. The appropriate procedure is to dilute the cells in accordance with the indicated guidelines.

Subculturing	Subculture by transferring an appropriate amount of the cell suspension into new cell culture flasks already containing fresh cell culture media.
Seeding density	Start cultures at 5 x 10^4 viable cells/ml.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	NFS-60 cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.
Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in a 50 ml centrifuge tube. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Examine cell morphology and confluency using a microscope. Finally, incubate the flasks at 37 degrees Celsius for at least 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

M_18-3: 16
M_4-2: 19.3/20.3
M_6-7: 11. Dez
M_3-2: 13/14
M_19-2: 11. Dez
M_7-1: 28/29
M_1-1: Okt 16
M_8-1: 15/16
M_2-1: Sep 16
M_15-3: 20.3/21.3
M_6-4: 15.03.2018
M_11-2: 17/18
M_1-2: 17
M_17-2: 13/15
M_12-1: 16/20
M_5-5: 14/15
M_X-1: 25/27
M_13-1: 13/14.2
Human D4/D8: -