Product sheet

Meth A sarcoma Cells | 400284

General information

Description	Meth A sarcoma cells have been established as in vitro cell line from the transplantable Meth A sarcoma tumor, which was induced in 1962 using 3-methylcholanthrene by Old et al. in Balb/c mice.
Organism	Mouse
Tissue	Skin
Disease	Fibrosarcoma
Synonyms	Meth A, Meth-A, Meth-A-sarkom

Characteristics

Age	Adult
Gender	Female
Morphology	Round cells
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	Meth A sarcoma (Cytion catalog number 400284)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

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Medium supplements	Supplement the medium with 10% FBS
Doubling time	28 to 30 hours
Subculturing	Allow cell aggregates to settle to the bottom of the flask, discard the supernatant medium, disperse the cells with gentle pipetting and dispense into new flasks. Resuspend cell suspension in the flask and take representative aliquot to count the cell number per ml. Dilute cell suspension to 1x10^5 cells/ml with fresh medium and transfer into new flasks.
Split ratio	A ratio of 1:4 to 1:8 is recommended
Seeding density	Start new cultures using 2 to 3 x 10^6 cells/ml. Once the cells have recovered from the freezing and thawing process after 1 to 2 passages, adjust the cell density to 1 x 10^6 cells/ml when splitting the cells.
Fluid renewal	2 to 3 times per week
Freezing recovery	About 53% of the initial cell number was collected after freezing.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

STR profile

Amelogenin: x,y