Product sheet

RT-112 Cells | 300324

General information

Description	This cell line was established by Dr. Carol Rigby, St. Paul?S Hospital, London as described by Benham et al. in 1976 from a human bladder carcinoma.
Organism	Human
Tissue	Bladder
Disease	Carcinoma
Synonyms	RT 112, RT112

Characteristics

Age	Unspecified
Gender	Female
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	RT-112 (Cytion catalog number 300324)
Biosafety level	1

Expression / Mutation

Protein expression	p53 positive, Cytokeratine (4),5,(6), 7, 8, 13, 17, 18, 19, Desmoplakin
Isoenzymes	Yes, in nude mice

Handling

Product sheet

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Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:4 to 1:8 is recommended
Seeding density	Start culture at 2 to 3 x 10^4 cells/cm^2 and continue with a seeding density of 1 x 10^4 cells/cm^2.
Fluid renewal	2 to 3 times per week
Freezing recovery	About 50% viable cells, which recover within 24 to 48 hours
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile	**Amelogenin**: x,y **CSF1PO**: 10,11 **D13S317**: 13,14 **D16S539**: 11,13 **D5S818**: 10,13 **D7S820**: 12,11 **TH01**: 7 **TPOX**: 8,11 **vWA**: 14,17 **D3S1358**: 15 **D21S11**: 27,30 **D18S51**: 15 **Penta E**: 12,16 **Penta D**: 10,11 **D8S1179**: 13,15 **FGA**: 23
HLA alleles	**A***: 26:01:01 **B***: 27:05:02 **C***: 01:02:01 **DRB1***: 01:01:01 **DQA1***: 01:01:01 **DQB1***: 05:01:01 **DPB1***: 01:01:01 **E**: 01:01:01