Product sheet

BEWO Cells | 300123

General information

Description	BeWo cells, a cell line derived from malignant gestational choriocarcinoma of the fetal male placenta, have become a widely used in vitro model for studying the placenta. The cell-cell fusion during the human trophoblast syncytialization phase during placental development is one of the most significant yet least understood events. Due to the difficulty of studying this process in a placenta in vivo, BeWo cells are utilized as a cell culture model to simulate in vivo syncytialization of the placental villous trophoblast. These cells exhibit an epithelial-like phenotype and are adherent. The b30 subclone of BeWo cells is particularly useful for studying nutrient uptake and transport due to its dense growth on permeable membranes. CK 7 and E-cadherin are molecular markers that are expressed by BeWo cells. VE-cadherin is found in BeWo cells and is enhanced upon treatment with forskolin. The cells also express keratin and are positive for G6PD, B isoenzyme. The karyotype of BeWo cells is modal number = 86, with a range of 71 to 178, and the stemline number is hypotetraploid. The karyotype is relatively stable within the stemline number. BeWo cells secrete various hormones, including human chorionic gonadotropin (hCG), human chorionic somatomammotropin (placental lactogen), and steroid hormones as estrone, estriol, and estradiol. However, the levels of β-hCG and estradiol secreted by BeWo cells are lower than those secreted by other choriocarcinoma-derived cell lines such as JEG-3. Upon Forskolin treatment, the secretion of β-hCG in BeWo cells increases to a level similar to that observed in the other choriocarcinoma-derived cell lines. Furthermore, Forskolin treatment also increases the progesterone levels secreted by BeWo cells. In summary, BeWo cells are a widely used in vitro model for studying placental development and the human trophoblast syncytialization process. They exhibit an epithelial-like phenotype, express various molecular markers, and secrete multiple hormones, including hCG, placental lactogen, and steroid hormones. Overall, BeWo cells are a valuable tool for investigating the complex processes involved in placental development.
Organism	Human
Tissue	Placenta
Disease	Choriocarcinoma
Metastatic site	Brain
Synonyms	BeWo, Be Wo, Be-Wo

Characteristics

Age	Fetus

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Gender	Male
Morphology	Epithelial-like
Growth properties	Adherent

Regulatory Data

Citation	BEWO (Cytion catalog number 300123)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0044

Biomolecular Data

Isoenzymes	G6PD, B
Virus susceptibility	Poliovirus 3, vesicular stomatitis (Indiana)
Reverse transcriptase	Negative
Products	Progesterone, human chorionic somatomammotropin (placental lactogen), estrogen, estrone, estriol, estradiol, keratin

Handling

Culture Medium	Ham's F12K Medium, w: 2.0 mM L-Glutamine, w: 2.0 mM Sodium pyruvate, w: 2.5 g/L NaHCO3 (Cytion article number 820608a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase

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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	A seeding density of 1 x 10^4 cells/cm^2 is recommended.
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Flask Coating	None
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.