

Capan-2 Cells | 300144**General information****Description**

The Capan-2 cell line is a human pancreatic adenocarcinoma cell line first isolated from the pancreatic tumor tissue of a 56-year-old Caucasian male. It was derived from the metastatic site in the liver, indicating its origin from a secondary tumor which makes it particularly valuable for research on metastatic processes and pancreatic cancer biology. The cells exhibit epithelial morphology and have been utilized extensively to study pancreatic cancer, drug resistance, and tumor biology.

Capan-2 cells are known to express a mutated form of the Kirsten rat sarcoma viral oncogene homolog (KRAS), a common mutation in pancreatic cancer, making them a robust model for studying KRAS-driven tumorigenesis. Additionally, they are characterized by the expression of tumor suppressor gene p53 mutations and have been observed to exhibit chromosomal instabilities, which are critical features relevant to cancer progression and treatment response. This cell line has been used in numerous studies, including those evaluating chemotherapeutic efficacy, exploring molecular pathways of cancer progression, and developing targeted therapy strategies.

Organism

Human

Tissue

Pancreas

Disease

Adenocarcinoma

Synonyms

CaPan-2, CAPAN-2, Capan 2, CAPAN 2, Capan2, CAPAN2

Characteristics**Age**

56 years

Gender

Male

Ethnicity

Caucasian

Morphology

Polygonal

Growth properties

Adherent, colonies

Regulatory Data**Citation**

Capan-2 (Cytion catalog number 300144)

Biosafety level

1

Capan-2 Cells | 300144**NCBI_TaxID** 9606**CellosaurusAccession** CVCL_0026**Biomolecular Data****Protein expression** P53 negative**Antigen expression** Blood Type B, Rh+**Isoenzymes** Me-2, 2, PGM3, 2, PGM1, 1, ES-D, 1, AK-1, 1, G6PD, B, GLO-1, 2, Phenotype Frequency Product: 0.0004**Tumorigenic** Yes, in nude mice. Forms well differentiated adenocarcinoma consistent with pancreatic carcinoma**Products** Mucin (apomucin, MUC-1, MUC-2)**Ploidy status** Aneuploid**Mutational profile** Capan-2 cells carry a heterozygous Kras mutation in codon12: GGT>GTT**Handling****Culture Medium** McCoys 5a, w: 3.0 g/L Glucose, w: stable Glutamine, w: 2.0 mM Sodium pyruvate, w: 2.2 g/L NaHCO₃ (Cytion article number 820200a)**Supplements** Supplement the medium with 10% FBS**Dissociation Reagent** Accutase**Doubling time** 45 to 60 hours**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

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Seeding density 1×10^4 cells/cm² will result in a confluent monolayer within 7 days.

Fluid renewal 2 to 3 times per week

Post-Thaw Recovery After thawing, plate the cells at 5×10^4 cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 48 hours.

Freeze medium As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere 37°C, 5% CO₂, humidified atmosphere.

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Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.