

**KATO-III Cells | 300381****General information****Description**

The KATO-III cell line is a human gastric carcinoma model derived from the metastatic site of a poorly differentiated adenocarcinoma. These cells are widely utilized in research focused on gastric cancer, particularly for studying the molecular mechanisms driving tumor progression, drug resistance, and metastasis. The KATO-III cells exhibit an aneuploid karyotype, characterized by multiple chromosomal abnormalities, which contributes to their aggressive cancer phenotype. They are notably p53 deficient, a feature often associated with increased tumorigenicity and altered responses to chemotherapy, making them a valuable tool for investigating the role of p53 in gastric cancer.

KATO-III cells grow in suspension and display a rounded morphology. They possess a high capacity for proliferation, making them suitable for various in vitro applications, including drug screening and cytotoxicity assays. These cells are also used in studies of cell signaling pathways, as their aberrant signaling is a hallmark of gastric cancer pathogenesis. Researchers often utilize KATO-III cells to explore the efficacy of novel therapeutic agents, particularly those targeting HER2, EGFR, and other relevant oncogenic pathways. This cell line is essential for advancing our understanding of gastric cancer biology and for developing targeted therapies aimed at improving patient outcomes.

**Organism** Human**Tissue** Stomach**Disease** Adenocarcinoma**Metastatic site** Pleural effusion**Synonyms** Kato III, Kato-III, KATO III, KATOIII, Katolll, KATO 3, JTC-28, Japanese Tissue Culture-28**Characteristics****Age** 57 years**Gender** Male**Ethnicity** Asian**Morphology** Spherical**Growth properties** Adherent/suspension**Regulatory Data**

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<b>Citation</b>	KATO-III (Cytion catalog number 300381)
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<b>Biosafety level</b>	1
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<b>NCBI_TaxID</b>	9606
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<b>CellosaurusAccession</b>	CVCL_0371
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## Biomolecular Data

<b>Protein expression</b>	p53 negative, CEA positive
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<b>Antigen expression</b>	Blood Type B, Rh+
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<b>Isoenzymes</b>	PGM3, 1, PGM1, 1, ES-D, 1, AK-1, 1, GLO-1, 2, G6PD, B, Phenotype Frequency Product: 0.0742
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<b>Tumorigenic</b>	Yes, in cheek pouches of anti thymocyte serum treated hamsters, not tumorigenic in nude mice
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<b>Karyotype</b>	The stemline chromosome number is hypotetraploid with the 2S component occurring at 6.2%. Nine markers were common to most S metaphases, four markers were less frequent. One (occasionally 2 copies) homogenous staining region (HSR) (t(11,HSR) was present in all metaphases examined, but no double minutes (DM) were detected (Sekiguchi 1978).
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## Handling

<b>Culture Medium</b>	Ham's F12, w: 1.0 mM stable Glutamine, w: 1.0 mM Sodium pyruvate, w: 1.1 g/L NaHCO <sub>3</sub> (Cytion article number 820600a)
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<b>Doubling time</b>	36 hours
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<b>Subculturing</b>	Gather the suspension cells in a 15 ml tube and gently wash the adherent cells with PBS lacking calcium and magnesium (use 3-5 ml for T25 flasks and 5-10 ml for T75 flasks). Apply Accutase (1-2 ml for T25 flasks, 2.5 ml for T75 flasks) ensuring full coverage of the cell layer. Allow the cells to incubate at room temperature for 10 minutes. Following incubation, combine and centrifuge both the suspension and adherent cells. After centrifugation, carefully resuspend the cell pellet and transfer the cell suspension into new flasks containing fresh medium.
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<b>Split ratio</b>	A ratio of 1:2 to 1:8 is recommended
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<b>Seeding density</b>	$2 \times 10^4$ cells/cm <sup>2</sup> will result in a confluent monolayer within 2 to 3 days.
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**Fluid renewal**      Every 3 to 5 days

**Freeze medium**      As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

## Quality Control & Molecular Analysis

**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

**STR profile**

**Amelogenin:** x,x  
**CSF1PO:** 7,11  
**D13S317:** 8,12  
**D16S539:** 10,12  
**D5S818:** 10,11  
**D7S820:** 8,12  
**TH01:** 7,9  
**TPOX:** 11  
**vWA:** 14,16  
**D3S1358:** 15,16  
**D21S11:** 30,31  
**D18S51:** 12  
**Penta E:** 13,18,19  
**Penta D:** 13,14  
**D8S1179:** 13,14  
**FGA:** 23,24

**HLA alleles**

**A\*:** '02:01:01, '02:07:01  
**B\*:** '15:01:01, '46:01:01  
**C\*:** '01:02:01, '03:03:01  
**DRB1\*:** '08:03:02, '15:01:01G  
**DQA1\*:** '01:02:01, '01:03:01  
**DQB1\*:** '06:01:01, '06:02:01  
**DPB1\*:** '02:01:02, '02:02:01  
**E:** '01:03:02