Product sheet

B-LCL-HROC06 Cells | 302065

General information

Organism	Human
Tissue	Peripheral blood
Disease	Carcinoma
Synonyms	Bc HROC06

Characteristics

Age	Age unspecified
Gender	Female
Ethnicity	Caucasian
Morphology	Round cells
Cell type	B lymphoblast
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	B-LCL-HROC06 (Cytion catalog number 302065)
Biosafety level	2
Depositor	M. Linnebacher

Expression / Mutation

Surface antigens	CD19

Handling

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Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% heat-inactivated FBS
Subculturing	Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	B-LCL-HROC06 cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.
STR profile	**Amelogenin**: x,x **CSF1PO**: 12, 12 **D13S317**: 11, 13 **D16S539**: 11, 12 **D5S818**: 9, 13 **D7S820**: 10, 12 **TH01**: 6, 9.3 **TPOX**: 8, 8 **vWA**: 16, 16 **D3S1358**: 15, 15 **D21S11**: 27, 30 **D18S51**: 18, 18 **Penta E**: 12, 12 **Penta D**: 10, 12 **D8S1179**: 14, 14 **FGA**: 22, 23