Product sheet

V79-4 Cells | 603371

General information

Description	The V79 cell line was developed by Ford and Yerganian in 1958 from lung tissue of a young male Chinese hamster, and was originally designated Strain V. Elkind renamed the line V-79 in 1958. The sub-clone V79-4 was isolated by E.H.Y. Chu In 1966, who had obtained the line from W. Sinclair.
Organism	Hamster
Tissue	Lung
Disease	Fibroblast
Synonyms	V-79-4, V 79-4

Characteristics

Age	Adult
Gender	Male
Morphology	Fibroblast-like
Growth properties	Monolayer, adherent

Identifiers / Biosafety / Citation

Citation	V79-4 (Cytion catalog number 603371)
Biosafety level	1

Expression / Mutation

Karyotype	Modal number = 22. Range = 20 to 23 Pseudodiploid. The rate of higher ploidies was 4%. Twelve to marker chromosomes were common to most cells. These include 1p-, 4q+, 4p+, t(6,?), 7q+ and seven to eight other small markers. Normal N2 and N3 were paired, N1, N5, N6 and N10 were single. Normal x and Y were absent, but a single xq- was present in every cell.

Handling

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Culture Medium	EMEM, w: 2 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: EBSS, w: 1 mM Sodium pyruvate, w: NEAA (Cytion article number 820100c)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	12 to 14 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:6 to 1:14 is recommended
Fluid renewal	1 to 2 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.