**Product sheet**

# SK-LMS-1 Cells | 300125

## General information

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| **Description** | SK-LMS-1 is a human leiomyosarcoma cell line that has been widely used for cancer research, particularly for studies investigating therapeutic agents targeting soft tissue sarcomas. Leiomyosarcoma is a type of malignant tumor that arises from smooth muscle tissues, and the SK-LMS-1 cell line models this disease effectively in vitro. These cells express the c-Met proto-oncogene, which plays a critical role in tumorigenesis, proliferation, and metastasis in many cancers, including leiomyosarcoma. The aberrant expression of c-Met in SK-LMS-1 makes it a valuable model for studying c-Met-targeted therapies.<br><br>One significant study involved the identification of a Met-binding peptide, Met-pep1, through phage display library screening. This peptide demonstrated specificity for the Met receptor and was capable of competing with hepatocyte growth factor (HGF) for receptor binding, inhibiting tumor cell proliferation. SK-LMS-1 cells treated with Met-pep1 showed decreased proliferation, suggesting that targeting c-Met with this peptide could have therapeutic potential. The internalization of the peptide by SK-LMS-1 cells after binding to c-Met further supports its potential as a diagnostic or therapeutic agent, particularly in nuclear imaging studies where tumor-associated activity was successfully visualized in vivo using SK-LMS-1 xenografts.<br><br>Additionally, SK-LMS-1 cells have been used to explore the effects of natural compounds such as Flavokawain B (FKB), a chalcone derived from the kava plant. FKB was found to induce G2/M cell cycle arrest and robust apoptosis in SK-LMS-1 cells, mediated by the upregulation of pro-apoptotic proteins like DR5, Bim, and Puma, and downregulation of the anti-apoptotic protein survivin. The combination of FKB with chemotherapeutic agents such as docetaxel and gemcitabine exhibited a synergistic effect, further inhibiting the growth of SK-LMS-1 cells. |
| **Organism** | Human |
| **Tissue** | Vulvar |
| **Disease** | Leiomyosarcoma |
| **Synonyms** | SKLMS-1, SKLMS1 |

## Characteristics

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| **Age** | 43 years |
| **Gender** | Female |
| **Ethnicity** | Caucasian |
| **Morphology** | Fibroblast-like |
| **Growth properties** | Adherent |

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## Regulatory Data

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| **Citation** | SK-LMS-1 (Cytion catalog number 300125) |
| **Biosafety level** | 1 |
| **NCBI_TaxID** | 9606 |
| **CellosaurusAccession** | CVCL_0628 |

## Biomolecular Data

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| **Antigen expression** | Blood Type O, Rh+ |
| **Isoenzymes** | Me-2, 2, PGM3, 1-2, PGM1, 1-2, ES-D, 1, AK-1, 1, GLO-1, 1-2, G6PD, B, Phenotype Frequency Product: 0.0027 |
| **Tumorigenic** | Yes, in nude mice. Forms leiomyosarcoma |
| **Karyotype** | (P12) hypotriploid to hypertriploid (+A2, +A3, +C, +D, +E, +F, +G, -A) with abnormalities including dicentrics, acrocentric fragments, breaks, secondary constrictions, minutes and large submetacentric markers |

## Handling

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| **Culture Medium** | DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a) |
| **Supplements** | Supplement the medium with 10% FBS |
| **Dissociation Reagent** | Accutase |
| **Subculturing** | Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium. |
| **Fluid renewal** | 2 to 3 times per week |

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| **Freeze medium** | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress. |
| **Thawing and Culturing Cells** | 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.<br>2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.<br>3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.<br>4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.<br>5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.<br>6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.<br>7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.<br>8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes. |
| **Incubation Atmosphere** | 37°C, 5% $CO_2$, humidified atmosphere. |
| **Shipping Conditions** | Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage. |
| **Storage Conditions** | For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen. |

## Quality Control & Molecular Analysis

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| **Sterility** | Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.<br><br>To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections. |