

## HCT-8 (HRT-18) Cells | 300210

### General information

#### Description

HCT-8 cells, also known as human ileocecal colorectal adenocarcinoma cells, are an epithelial cell line originally derived from a 67-year-old Caucasian male patient with ileocecal adenocarcinoma. The HCT-8 cell line was established in the late 1960s and is widely utilized in cancer research, particularly in the study of colorectal cancer pathogenesis, metastasis, and treatment response.

Morphologically, HCT-8 cells are epithelial-like and exhibit a monolayer growth pattern with a polygonal shape. They possess the capability to grow in both adherent and semi-suspended cultures, which is characteristic of some transitional stages of cancer cell metastasis. This feature makes them particularly useful for studies related to cancer cell invasion and migration.

Genotypically, HCT-8 cells are hypertriploid, containing several chromosomal aberrations common in colorectal carcinomas, including mutations and deletions which are relevant to cancer progression and resistance mechanisms. This genetic profile supports their use in oncological studies, especially those focusing on genetic pathways involved in tumorigenesis and drug resistance.

Research utilizing HCT-8 cells has contributed significantly to the understanding of colorectal cancer biology, including the elucidation of molecular pathways involved in cancer cell proliferation, apoptosis, and chemoresistance. The cell line continues to be a critical model for investigating the efficacy of new therapeutic agents and for exploring the molecular mechanisms underlying colorectal cancer.

**Organism** Human

**Tissue** Rectum

**Disease** Adenocarcinoma

**Synonyms** HCT 8, HCT8

### Characteristics

**Age** 67 years

**Gender** Male

**Morphology** Epithelial-like

**Growth properties** Adherent

### Regulatory Data

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<b>Citation</b>	HCT-8 (Cytion catalog number 300210)
<b>Biosafety level</b>	1
<b>NCBI_TaxID</b>	9606
<b>CellosaurusAccession</b>	CVCL_2478

### Biomolecular Data

<b>Antigen expression</b>	CDx (+/-), CDy (-),
<b>Isoenzymes</b>	AK-1, 1, ES-D, 1-2, GLO-1, 2, G6PD, B, PGM1, 1, PGM3, 1, Me-2, 1
<b>Tumorigenic</b>	In nude mice
<b>Viruses</b>	Reverse Transcriptase negative
<b>Products</b>	Carcinoembryonic antigen (CEA) 0.5 ng/10 exp6 cells/10 days, alkaline phosphatase, keratin
<b>Mutational profile</b>	HRT-18 cells carry a mutation in codon 13 of Kras gene: GGC(Wt Gly) >GAC(Asp)

### Handling

<b>Culture Medium</b>	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
<b>Supplements</b>	Supplement the medium with 10% FBS
<b>Dissociation Reagent</b>	Accutase
<b>Doubling time</b>	15 hours
<b>Subculturing</b>	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

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**Seeding density** 2 to 4 x 10<sup>4</sup> cells/cm<sup>2</sup>

**Fluid renewal** 2 to 3 times per week

**Post-Thaw Recovery** Fast

**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

**Incubation Atmosphere** 37°C, 5% CO<sub>2</sub>, humidified atmosphere.

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### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately  $-78^{\circ}\text{C}$  throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about  $-150$  to  $-196^{\circ}\text{C}$ . Storage at  $-80^{\circ}\text{C}$  is acceptable only as a short interim step before transfer to liquid nitrogen.

## Quality Control & Molecular Analysis

### Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.