Product sheet

SW-1353 Cells | 300440

General information

Description	This cell line was established in 1977 by A. Leibovitz. The initial culture medium was Leibovitz medium (L-15) containing cortisone and insulin plus 10% fetal bovine serum and antibiotics.
Organism	Human
Tissue	Bone, right humerus
Disease	Chondrosarcoma (Grade II)
Synonyms	SW1353, SW 1353

Characteristics

Age	72 years
Gender	Female
Ethnicity	Caucasian
Morphology	Fibroblast-like
Growth properties	Monolayer, adherent

Identifiers / Biosafety / Citation

Citation	SW-1353 (Cytion catalog number 300440)
Biosafety level	1

Expression / Mutation

Antigen expression	Antigen Expression: Blood type O-
Isoenzymes	G6PD, B, PGM1, 1, PGM3, 2, ES-D, 2, AK-1, 1, GLO-1, 2, Phenotype Frequency Product: 0.00009
Karyotype	hyperdiploid 47,xx, +7, Except for the trisomic N7 no other chromosome markers are evident

Handling

Culture Medium	DMEM:Ham's F12, w: 3.1 g/L Glucose, w: 1.6 mM L-Glutamine, w: 15 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:3 is recommended
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile	**CSF1PO**: 12 **D13S317**: 12,13 **D16S539**: 11,12 **D5S818**: 10,11 **D7S820**: 9,11 **TH01**: 6,9 **TPOX**: 8,11 **vWA**: 16,17 **D3S1358**: 15,18 **D21S11**: 30,32.2 **D18S51**: 13,17 **Penta E**: 12,14 **Penta D**: 10,11 **D8S1179**: 10,11 **FGA**: 22,23
HLA alleles	**A***: 24:02:01, 29:02:01 **B***: 44:02:01, 44:03:01 **C***: 02:02:02, 16:01:01 **DRB1***: 07:01:01, 13:01:01 **DQA1***: 01:03:01, 02:01:01 **DQB1***: 02:02:01, 06:03:01 **DPB1***: 02:01:02, 04:01:01 **E**: 01:01:01, 01:03