Product sheet

KYSE-30 Cells | 305094

General information

Description	This cell line was established from the mucosal surface of a well differentiated invasive esophageal squamous cell carcinoma from a 64-year-old Japanese male prior to treatment. The cell line was used for heterotransplanting tumours to athymic mice. The cells have a p53 mutation at the splice acceptor site and amplification of cERB B, MYC and CYCLIN D1.
Organism	Human
Tissue	Esophageal Squamous Epithelium
Disease	Esophageal squamous cell carcinoma
Synonyms	Kyse-30, KYSE 30, KYSE30, Kyse30, KYSE0030

Characteristics

Age	64 years
Gender	Male
Ethnicity	Asian
Morphology	Epithelial-Like, With Long Pseudopod
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	KYSE-30 (Cytion catalog number 305094)
Biosafety level	1

Expression / Mutation

Handling

Culture Medium	Please mix Ham's F12 and RPMI 1640 in a 50:50 ratio (Cytion article numbers 820600a and 820702a)

Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Doubling time	20 to 30 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1: 3 to 1: 5
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	KYSE-30 cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Product sheet

KYSE-30 Cells | 305094

STR profile

Amelogenin: x,x
CSF1PO: 10
D13S317: 9
D16S539: 10,12
D5S818: 11
D7S820: 11,11.3
TH01: 9
TPOX: 9
vWA: 16,18,19
D3S1358: 15,16
D21S11: 28
D18S51: 14
Penta E: 13
Penta D: 12
D8S1179: 12,15
FGA: 24
D6S1043: 11,20
D2S1338: 23
D12S391: 17,19
D19S433: 14.2,15.2