Product sheet

WEHI-3B Cells | 400376

General information

Description	The growth of WEHI-3 is inhibited by 4 ng/ml LPS and blocked by higher concentrations. Dextran sulfate at 30 to 40 microgram/ml also inhibits growth. Latex beads are phagocytized but are not toxic. Zymosan and BCG are phagocytized and block growth. The cells exhibit only weak effector activity in antibody dependent cell mediated cytotoxicity.
Organism	Mouse
Tissue	Peripheral blood
Disease	Leukemia
Synonyms	WEHI-3b, Wehi-3B, WEHI 3B, WEHI3B

Characteristics

Cell type	Myelomonocyte
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	WEHI-3B (Cytion catalog number 400376)
Biosafety level	2

Expression / Mutation

Receptors expressed	Immunoglobulin (Fc), complement (C3)
Viruses	Ectromelia virus (mousepox) negative
Products	Lysozyme, granulocyte colony stimulating activity (G-CSA), interleukin-3 (interleukin 3, IL-3)

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Product sheet

WEHI-3B Cells | 400376

Medium supplements	Supplement the medium with 10% FBS
Subculturing	Cultures can be maintained by addition or replacement of fresh medium. Start cultures at 2 x 10^5 cells/ml and maintain between 1 x 10^5 and 1x10^6 cells/ml. Adherent cells can be recovered by scraping.
Seeding density	1 x 10^5 cells/ml
Fluid renewal	2 to 3 times per week
Freezing recovery	After thawing, allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.
STR profile	**Amelogenin**: x,x **M_18-3**: 17,20 **M_4-2**: 21.3 **M_6-7**: 12 **M_3-2**: 14 **M_19-2**: 13 **M_7-1**: 25.2,26.2 **M_1-1**: 15,16 **M_8-1**: 13 **M_2-1**: 16 **M_15-3**: 22.3 **M_6-4**: 18 **M_11-2**: 18,19 **M_1-2**: 17 **M_17-2**: 18 **M_12-1**: 16,17 **M_5-5**: 14,17 **M_X-1**: 26 **M_13-1**: 15,2 **Human D4/D8**: -