Product sheet

WEHI-3B Cells | 400376

General information

Description	The growth of WEHI-3 is inhibited by 4 ng/ml LPS and blocked by higher concentrations. Dextran sulfate at 30 to 40 microgram/ml also inhibits growth. Latex beads are phagocytized but are not toxic. Zymosan and BCG are phagocytized and block growth. The cells exhibit only weak effector activity in antibody dependent cell mediated cytotoxicity.
Organism	Mouse
Tissue	Peripheral blood
Disease	Leukemia
Synonyms	WEHI-3b, Wehi-3B, WEHI 3B, WEHI3B

Characteristics

Cell type	Myelomonocyte
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	WEHI-3B (Cytion catalog number 400376)
Biosafety level	2

Expression / Mutation

Receptors expressed	Immunoglobulin (Fc), complement (C3)
Viruses	Ectromelia virus (mousepox) negative
Products	Lysozyme, granulocyte colony stimulating activity (G-CSA), interleukin-3 (interleukin 3, IL-3)

Handling

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)

Medium supplements	Supplement the medium with 10% FBS
Passaging solution	The utilization of a passaging solution is not necessary when passaging cells that are cultured in suspension. The appropriate procedure is to dilute the cells in accordance with the indicated guidelines.
Subculturing	Cultures can be maintained by addition or replacement of fresh medium. Start cultures at 2 x 10^5 cells/ml and maintain between 1 x 10^5 and 1x10^6 cells/ml. Adherent cells can be recovered by scraping.
Seeding density	1 x 10^5 cells/ml
Fluid renewal	2 to 3 times per week
Freezing recovery	After thawing, allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	WEHI-3B cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.
Handling of proliferating cultures	One or two cell culture flasks come filled with cell culture medium. Collect the entire medium in a 50 ml centrifuge tube. Spin down the collected medium at 300 x g for 3 minutes to collect the cells which may have detached during transit. If a cell pellet is visible, resuspend the cells in 5 ml of cell culture medium and transfer to a T25 cell culture flask. Carefully add 5 ml of cell culture medium to each T25 cell culture flask. Examine cell morphology and confluency using a microscope. Finally, incubate the flasks at 37 degrees Celsius for at least 24 hours.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Amelogenin: x,x
M_18-3: 17/20
M_4-2: 21,3
M_6-7: 12
M_3-2: 14
M_19-2: 13
M_7-1: 25,2/26,2
M_1-1: 15/16
M_8-1: 13
M_2-1: 16
M_15-3: 22,3
M_6-4: 18
M_11-2: 18/19
M_1-2: 17
M_17-2: 18
M_12-1: 16/17
M_5-5: 14/17
M_X-1: 26
M_13-1: 15,2
Human D4/D8: -