Product sheet

AS-30D Cells | 500116

General information

Description	Established in vitro from the AS-30D tumor ascites.
Organism	Rat
Tissue	Liver
Disease	Hepatocellular carcinoma
Synonyms	A-S-30D, AS30D

Characteristics

Age	16 months
Gender	Female
Morphology	Round cells, loosely adherent, floating
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	AS-30D (Cytion catalog number 500116)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes, in Sprague-Dawley rats
Viruses	RAP-test: Negative.
Karyotype	Hypodiploid rat karyotype with 12% tetraploidy, 38 (35-41).

Handling

Product sheet

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Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Doubling time	26 hours
Subculturing	Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.
Split ratio	A ratio of 1:4 to 1:5 is recommended
Seeding density	A seeding density of 1 x 10^6 cells/ml is recommended.
Fluid renewal	Every 3 to 5 days
Freezing recovery	After thawing, allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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STR profile

Rat_D2Wox37: 150,152
Rat_D19Wox11: 228
Rat_D10Wox8: 266
Rat_D4Wox7: 153,157
Rat_D2Wox27: 211
Rat_D5Rat33: 122,124,128
Rat_D10Wox11: 156
Rat_D1Wox23: 210,214
Rat_D12Wox1: 410
Rat_D6Wox2: 104
Rat_D8Wox7: 182
Rat_D6Cebr1: 225
SRY: x,Y