Product sheet

AS-30D Cells | 500116

General information

Description	Established in vitro from the AS-30D tumor ascites.
Organism	Rat
Tissue	Liver
Disease	Hepatocellular carcinoma
Synonyms	A-S-30D, AS30D

Characteristics

Age	16 months
Gender	Female
Morphology	Round cells, loosely adherent, floating
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	AS-30D (Cytion catalog number 500116)
Biosafety level	1

Expression / Mutation

Tumorigenic	Yes, in Sprague-Dawley rats
Viruses	RAP-test: Negative.
Karyotype	Hypodiploid rat karyotype with 12% tetraploidy, 38 (35-41).

Handling

AS-30D Cells | 500116

Culture Medium	RPMI 1640, w: 2.1 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Medium supplements	Supplement the medium with 10% FBS
Doubling time	26 hours
Subculturing	Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.
Split ratio	A ratio of 1:4 to 1:5 is recommended
Seeding density	A seeding density of 1 x 10^6 cells/ml is recommended.
Fluid renewal	Every 3 to 5 days
Freezing recovery	After thawing, allow the cells to recover from the freezing process for at least 24 hours.
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	AS-30D cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Product sheet

AS-30D Cells | 500116

STR profile

Rat_D2Wox37: 150,152
Rat_D19Wox11: 228
Rat_D10Wox8: 266
Rat_D4Wox7: 153,157
Rat_D2Wox27: 211
Rat_D5Rat33: 122,124,128
Rat_D10Wox11: 156
Rat_D1Wox23: 210,214
Rat_D12Wox1: 410
Rat_D6Wox2: 104
Rat_D8Wox7: 182
Rat_D6Cebr1: 225
SRY: x,Y