Product sheet

NCI-H3122 Cells | 300484

General information

Description	The NCI-H3122 cell line is derived from non-small-cell lung cancer (NSCLC) and is characterized by the presence of the EML4-ALK fusion gene, which results from a chromosomal translocation between echinoderm microtubule-associated protein-like 4 (EML4) and anaplastic lymphoma kinase (ALK). This fusion drives oncogenic signaling and makes NCI-H3122 cells highly dependent on ALK signaling for survival, known as "ALK-addicted." NCI-H3122 has become a key model for studying targeted therapies, particularly for ALK inhibitors like crizotinib. Studies have shown that NCI-H3122 cells are sensitive to crizotinib, which inhibits ALK phosphorylation and its downstream targets such as the AKT and ERK pathways. However, resistance to crizotinib often develops, typically due to alternative signaling pathways like the activation of the epidermal growth factor receptor (EGFR). This resistance mechanism has been confirmed in NCI-H3122 resistant variants, where increased EGFR phosphorylation was observed, and dual inhibition of ALK and EGFR using crizotinib and EGFR inhibitors such as afatinib or erlotinib was shown to overcome the resistance. NCI-H3122 is frequently used to explore combination therapies aimed at preventing or reversing drug resistance. For instance, targeting both ALK and EGFR pathways has been a successful strategy in preclinical models, and this dual inhibition has been suggested as a potential therapeutic approach for ALK-positive, crizotinib-resistant NSCLC patients.
Organism	Human
Tissue	Lung
Disease	Adenocarcinoma
Synonyms	NCI-H3122, H-3122, NCIH3122

Characteristics

Gender	Male
Ethnicity	Caucasian
Growth properties	Adherent

Regulatory Data

Citation	NCI-H3122 (Cytion catalog number 300484)
Biosafety level	1

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NCBI_TaxID	9606
CellosaurusAccession	CVCL_5160

Biomolecular Data

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.