Product sheet

Sp2/0-Ag14 Cells | 400481

General information

Description	The line was formed by fusing Balb/c spleen cells (from mouse immunized with sheep RBCs) with the P3x63Ag8 myeloma cell line. The cells do not secrete immunoglobulin, are resistant to 8-azaguanine at 20 µg/ml and to 6-thioguanine at 30 µM, and they are HAT sensitive.. Sp2/0-Ag14 cells can be used as fusion partners for B cells in the production of hybridomas.
Organism	Mouse
Tissue	Blood
Disease	B cell hybridoma
Synonyms	SP2/0-Ag14, SP2/0-AG14, SP2/0-ag14, Sp2/O-Ag14, SP2/O-Ag14, Sp2/0-Ag-14, SP2-0-Ag14, SP2/0 Ag-14, SP-2/0-AG14, Sp 2/0-Ag 14, Sp2/0, SP2/0, Sp2/O, SP2/O, SP-2, SP2, GM03569, GM3569, GM03569B, GM3569B, GM03569D

Characteristics

Morphology	Round cells
Growth properties	Suspension

Identifiers / Biosafety / Citation

Citation	Sp2/0-Ag14 (Cytion catalog number 400481)
Biosafety level	1
Depositor	T. Lindl

Expression / Mutation

Antigen expression	H-2d
Viruses	Tested and found negative for ectromelia virus (mousepox).

Handling

Sp2/0-Ag14 Cells | 400481

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS
Subculturing	Collect medium with floating cells in a microcentrifuge tube. Rinse the adherent cells using PBS without calcium and magnesium (3-5 ml PBS for T25, 5-10ml for T75 cell culture flasks). Add Accutase (1-2ml per T25, 2.5ml per T75 cell culture flask), the cell sheet must be covered completely. Incubate at 37 degree Celsius for 10 minutes. Combine the floating cells and the detached cells in one tube, centrifuge at 300xg for 3min. Carefully resuspend the cells in fresh medium and dispense into new flasks which contain fresh medium.
Seeding density	Maintain cell density between 5 x 10^4 and 5 x 10^6 viable cells/ml.
Fluid renewal	2 to 3 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	Sp2/0-Ag14 cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

Sp2/0-Ag14 Cells | 400481

STR profile

Amelogenin: x,x
M_18-3: 17,18,19,20
M_4-2: 21.3
M_6-7: 12,13
M_3-2: 13,14,15
M_19-2: 12,13
M_7-1: 24.2,25.2
M_1-1: 16,17,19
M_8-1: 13
M_2-1: 15,16
M_15-3: 21.3,23.3
M_6-4: 18,19
M_11-2: 17
M_1-2: 16,17
M_17-2: 16
M_12-1: 15,16
M_5-5: 14,15
M_X-1: 25,26
M_13-1: 16.2,17.2,18.2
Human D4/D8: -