**Product sheet**

# L-540 Cells | 300201

## General information

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| **Description** | L-540 is a human Hodgkin's lymphoma cell line derived from a patient with this form of cancer. This cell line is extensively utilized in research focused on the molecular and cellular mechanisms underlying Hodgkin's lymphoma, a malignancy originating from B lymphocytes. L-540 cells exhibit the characteristic Reed-Sternberg cells, which are a hallmark of Hodgkin's lymphoma and critical for diagnosing this disease. The presence of these multinucleated giant cells makes L-540 an invaluable model for studying the pathophysiology of Hodgkin's lymphoma and for screening potential therapeutic agents targeting these malignant cells.<br><br>One of the notable features of L-540 is its expression of CD30, a member of the tumor necrosis factor receptor family, which is often overexpressed in Hodgkin's lymphoma cells. This makes L-540 an excellent model for investigating CD30-targeted therapies, such as antibody-drug conjugates. Additionally, L-540 cells have been used to study the effects of various chemotherapeutic agents and to explore the mechanisms of drug resistance in lymphoma. The cell line's ability to form tumors in immunocompromised mice further enhances its utility in preclinical studies aimed at evaluating the efficacy of new treatments for Hodgkin's lymphoma. |
| **Organism** | Human |
| **Tissue** | Bone marrow |
| **Disease** | Hodgkin lymphoma |
| **Synonyms** | L 540, L540 |

## Characteristics

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| **Age** | 20 years |
| **Gender** | Female |
| **Ethnicity** | European |
| **Morphology** | Round cells |
| **Growth properties** | Suspension |

## Regulatory Data

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| **Citation** | L-540 (Cytion catalog number 300201) |
| **Biosafety level** | 1 |

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| **NCBI_TaxID** | 9606 |
| **CellosaurusAccession** | CVCL_1362 |

## Biomolecular Data

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| **Viruses** | Transformed by EBV |

## Handling

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| **Culture Medium** | RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a) |
| **Supplements** | Supplement the medium with 10% FBS |
| **Subculturing** | Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x $10^5$ cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation. |
| **Fluid renewal** | 3 times per week |
| **Freeze medium** | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress. |

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| **Thawing and Culturing Cells** | 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.<br>2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.<br>3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.<br>4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.<br>5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.<br>6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.<br>7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.<br>8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes. |
| **Incubation Atmosphere** | 37°C, 5% $CO_2$, humidified atmosphere. |
| **Shipping Conditions** | Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage. |
| **Storage Conditions** | For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen. |

## Quality Control & Molecular Analysis

**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.