

HEK293-CLDN6 Cells | 305985

General information

Description

Disclaimer: The prices displayed for cell lines are exclusively for academic/not-for-profit customers. For commercial entities the price is approximately €6,250.

If you represent a commercial entity or are unsure which category applies, please [contact us](#).

HEK293-CLDN6 cells are human embryonic kidney 293 (HEK293) cells engineered to stably express human claudin-6 (CLDN6), a tight junction-associated transmembrane protein belonging to the claudin family. CLDN6 is normally expressed during embryonic and fetal development but is largely absent in most healthy adult tissues, making it an attractive oncofetal antigen for targeted cancer therapy. Aberrant re-expression of CLDN6 has been identified in multiple malignancies, including ovarian cancer, testicular germ cell tumors, endometrial cancer, gastric cancer, and certain sarcomas. Stable HEK293-CLDN6 models provide a controlled system for studying CLDN6 biology and evaluating CLDN6-directed therapeutic approaches.

HEK293-CLDN6 cells are widely used in oncology research and drug development for characterization of monoclonal antibodies, antibody-drug conjugates, bispecific antibodies, CAR-T cell therapies, and other engineered immune cell platforms targeting CLDN6. The stable recombinant expression system supports quantitative assessment of antigen binding affinity, receptor density, antibody internalization, epitope specificity, and target-dependent cytotoxicity. These cells are also commonly applied in flow cytometry assay development, reporter assays, high-throughput therapeutic screening, and validation of CLDN6-targeted imaging agents. Because HEK293 cells exhibit high transfection efficiency and robust protein expression, they provide a reliable platform for recombinant membrane protein production and standardized assay generation.

Organism Human

Tissue Fetal Kidney

Characteristics

Age Fetus

Gender Female

Morphology Epithelial-like

Growth properties Monolayer, adherent

Regulatory Data

Citation HEK293-CLDN6 (Cytion catalog number 305985)

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Biosafety level 1**NCBI_TaxID** 9606**Biomolecular Data****Receptors expressed** CLDN6**Handling****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS, 1 mM sodium pyruvate, 10 mM HEPES, 1% NEAA. Add Geneticin (G418-Sulfat) to achieve a final concentration of 1 mg/mL.**Dissociation Reagent** Trypsin-EDTA**Subculturing** For routine adherent cell culture: Aspirate the old culture medium from the adherent cells, and wash them with PBS to remove any remaining medium. After aspirating the PBS, add the appropriate volume of Trypsin/EDTA solution based on the culture vessel size (e.g., 1 ml for a T25 flask, 3 ml for a T75 flask) and incubate at room temperature or 37°C until the cells detach (5-10 minutes). Monitor detachment under a microscope, and gently tap the vessel if necessary to release the cells. Once detached, add complete medium to inactivate the Trypsin/EDTA, gently resuspend the cells, and transfer an aliquot of the cell suspension into a new culture vessel containing fresh medium. Place the vessel in an incubator set to 37°C with 5% CO₂, and change the medium every 2-3 days.**Fluid renewal** 2 to 3 times per week**Post-Thaw Recovery** After thawing, split the cells at a ratio of 1:2 to 1:3 in T25 flasks and allow the cells to recover from the freezing process and to adhere (for adherent cultures) for at least 24 hours.**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.