Product sheet

A549 Cells | 300114

General information

Description	A549 cells, derived from lung adenocarcinoma tissue, are a primary model used in cancer research, particularly in biomedical laboratories focusing on lung-related cancers. A549 cells are commonly used as an in vitro model for studying lung cancer biology, drug screening, and the effects of toxic compounds. In toxicology research, A549 cells offer a controlled experimental model that enables scientists to explore the mechanisms underlying toxic effects and cellular responses. By understanding these mechanisms, researchers can better assess the safety of substances and potentially mitigate their harmful effects. A549 carcinoma cells have been extensively used as an in vitro model to study lung cancer pathogenesis and as an alternative tissue culture model for various pulmonary-related research studies in biomedical laboratories. These cells maintain the characteristics of type II alveolar epithelial cells and are used to examine the epithelial responses to various infections and inflammatory stimuli, including lung inflammation. Furthermore, the human cell line A549 serves as a valuable tool in the development of specific antibodies targeting lung cancer-related proteins or markers. By exposing these cells to substances of interest, researchers can investigate how they affect cell viability, proliferation, apoptosis, and other cellular processes. This information aids in the identification of potential therapeutic targets and the development of novel treatments for lung cancer. In summary, A549 carcinoma cells are pivotal in cancer research, especially concerning lung-related cancers, serving as an in vitro model for cancer and toxicology research, developing effective treatments, and drug screening.
Organism	Human
Tissue	Lung
Disease	Carcinoma
Synonyms	A 549, A-549, NCI-A549, hA54

Characteristics

Age	58 years
Gender	Male
Ethnicity	Caucasian
Morphology	Epithelial-like

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Growth properties	Adherent

Regulatory Data

Citation	A549 (Cytion catalog number 300114)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0023

Biomolecular Data

Protein expression	P53 positive
Isoenzymes	G6PD, type B
Reverse transcriptase	Negative
Karyotype	A549 cells have the modal chromosome number n2, with some cells with 64 chromosomes.

Handling

Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Doubling time	28 hours

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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.