

143B Cells | 305232

General information

Description

The 143B cell line is a human osteosarcoma cell line derived from a bone tumor. It is often used in cancer research due to its high metastatic potential and ability to form osteosarcoma tumors in vivo. These cells exhibit several key characteristics of osteosarcoma, including the expression of osteoblastic markers and the ability to produce osteoid. The 143B cell line is especially valuable for studying the mechanisms underlying bone cancer progression and metastasis, as well as for testing potential therapeutic agents aimed at treating osteosarcoma.

143B cells are known for their rapid growth and high transfection efficiency, making them suitable for genetic manipulation and various experimental applications. These cells have been utilized in studies investigating the role of specific genes and signaling pathways in osteosarcoma development and resistance to chemotherapy. Additionally, the 143B cell line serves as a model for exploring the interactions between osteosarcoma cells and the bone microenvironment, providing insights into the complex biology of bone tumors.

Organism Human

Tissue Bone, right femur

Disease Osteosarcoma

Synonyms 143b, 143 B, 143B TK-, 143B.TK-, 143BTK-, 143TK-, HOS-143B, HOS-143b, GM05887, GM05887A

Characteristics

Age 13 years

Gender Female

Ethnicity Caucasian

Growth properties Adherent

Regulatory Data

Citation 143B (Cytion catalog number 305232)

Biosafety level 1

NCBI_TaxID 9606

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CellosaurusAccession CVCL_2270

Biomolecular Data**Handling**

Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO ₃ , w: EBSS (Cytion article number 820100a)
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Supplements	Supplement the medium with 10% FBS and 1% NEAA
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Dissociation Reagent	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Seeding density	2×10^4 cells/cm ²
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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.