Product sheet

MDA-MB-468 Cells | 300279

General information

Description	The MDA-MB-468 cell line is a well-established human breast cancer cell line derived from the pleural effusion of an adult patient with metastatic adenocarcinoma. These cells are characterized by their epithelial morphology and are known for their high degree of aneuploidy. MDA-MB-468 cells are estrogen receptor-negative (ER-) and are often used as a model to study triple-negative breast cancer (TNBC), a subtype of breast cancer that lacks estrogen receptor (ER), progesterone receptor (PR), and HER2/neu expression. This makes MDA-MB-468 a critical tool for research into cancers that do not respond to hormonal therapy or HER2-targeted treatments. Genetically, MDA-MB-468 cells exhibit mutations in the TP53 gene, which is a common occurrence in various forms of cancer and plays a significant role in cell cycle regulation and apoptosis. The cell line also shows amplification of the epidermal growth factor receptor (EGFR) gene, contributing to its utility in studying the EGFR signaling pathway and its implications in cancer progression and treatment resistance. Researchers frequently utilize MDA-MB-468 cells to investigate mechanisms of drug resistance, test new therapeutic agents, and explore the molecular biology of aggressive breast cancers. In addition to their genetic and phenotypic characteristics, MDA-MB-468 cells are known for their ability to form xenografts in immunocompromised mice, making them a valuable model for in vivo studies of tumor growth and metastasis. This cell line's responsiveness to various chemotherapeutic agents and targeted therapies is extensively studied to develop effective treatment strategies for TNBC. Overall, the MDA-MB-468 cell line is a crucial resource for advancing breast cancer research, particularly in the context of triple-negative and EGFR-positive malignancies.
Organism	Human
Tissue	Breast
Disease	Adenocarcinoma
Metastatic site	Pleural effusion
Synonyms	MDA-MB 468, MDA-MB468, MDAMB468, MDA-468, MDA468, MB468, MD Anderson-Metastatic Breast-468

Characteristics

Age	51 years
Gender	Female
Ethnicity	African
Morphology	Epithelial

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Growth properties	Adherent

Regulatory Data

Citation	MDA-MB-468 (Cytion catalog number 300279)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0419

Biomolecular Data

Handling

Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.