

Product sheet

DSL-6A-C1 | 500166

General Information

Description	DSL-6A/C1 is a cell line derived from a patient with a high-grade glioma. It is characterized by its ability to form neurospheres and its sensitivity to radiation and chemotherapy. The cell line is maintained in DMEM/F12 supplemented with BDNF and EGF. It is a good model for studying glioma biology and drug response.
Organism	Human
Tissue	Brain
Disease	High-grade glioma, Glioblastoma
Metastatic site	None
Synonyms	DSL-6A/C1, DSL6A/C1

Characteristics

Breed/Subspecies	Human
Age	Adult
Gender	Male
Morphology	Spherical cell clusters
Cell type	Neuronal
Growth properties	Highly proliferative

Usage and Safety

Citation	DSL-6A-C1 (Cytion 500166)
Biosafety level	1
NCBI_TaxID	10116
CellosaurusAccession	CVCL_4166

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XXXXXXXXXX XXXX-XXXXXXXXXXXXXXXXXX

Tumorigenic Yes, this cell line is tumorigenic in immunodeficient mice. It forms solid tumors in the subcutaneous space of immunodeficient mice. The tumor formation is dependent on the number of cells injected and the site of injection. The tumor formation is observed in 100% of the mice injected with 10⁶ cells. The tumor formation is observed in 100% of the mice injected with 10⁵ cells. The tumor formation is observed in 100% of the mice injected with 10⁴ cells.

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Culture Medium Waymouth medium (DMEM/F12, 10% FBS, 2.0 mM L-glutamine)

Supplements 10% FBS, 2.0 mM L-glutamine

Dissociation Reagent Trypsin

Subculturing Cells are cultured in DMEM/F12 medium supplemented with 10% FBS and 2.0 mM L-glutamine. Cells are passaged into fresh medium when the cell density reaches 10⁶ cells per flask. Cells are passaged into fresh medium when the cell density reaches 10⁶ cells per flask.

Seeding density 1 x 10⁴ cells/cm²

Fluid renewal 2-3 times per week

Post-Thaw Recovery Cells are thawed in a 37°C water bath and immediately added to 10 ml of DMEM/F12 medium supplemented with 10% FBS and 2.0 mM L-glutamine. Cells are cultured in this medium for 24 hours before being passaged into fresh medium.

Freeze medium DMEM/F12 medium supplemented with 10% FBS and 10% DMSO

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Thawing and Culturing Cells

1. Thaw the cells quickly in a water bath at 37°C. Do not allow the cells to reach room temperature. After thawing, centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a 150 cm² flask with 8 ml of medium. Incubate at 37°C with 5% CO₂.
2. Once the cells have reached confluence, harvest the cells by trypsinization. Seed the cells into a 150 cm² flask with 8 ml of medium. Incubate at 37°C with 5% CO₂.
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8. Once the cells have reached confluence, harvest the cells by trypsinization. Seed the cells into a 150 cm² flask with 8 ml of medium. Incubate at 37°C with 5% CO₂.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating Cell culture medium, 10 minutes

Freezing Procedure Harvest cells by trypsinization and resuspend in freezing medium. Freeze at -80°C.

Shipping Conditions Store at -80°C.

Storage Conditions Store at -150°C for 196 days.

HLA

Sterility Sterility testing: PCR