

Product sheet

HOS | 300449

General Information

Description	HOS (ATCC CCL-TE85) is a continuous cell line derived from a 13-year-old male patient with osteosarcoma. HOS cells are derived from the primary tumor and are characterized by their ability to form osteoid matrix in culture. HOS cells are a derivative of the 143B cell line.
Organism	Human
Tissue	Osteosarcoma
Disease	Osteosarcoma

Characteristics

Age	13 years
Gender	Male
Ethnicity	White
Morphology	Epithelial cells, adherent, epithelial morphology
Growth properties	Highly proliferative, anchorage dependent

Identification and Safety

Citation	HOS (ATCC CCL-TE85) Cytion 300449
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0312

Enzymes and Markers

Isoenzymes	G6PD, B
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HEK293T HOS | 300449

HEK293T

Culture Medium EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO₃, w: EBSS (Cytion Cytion 820100a)

Supplements Cytion Cytion 10% FBS 1% NEAA

Dissociation Reagent Cytion Cytion

Subculturing HEK293T cells are cultured in EMEM supplemented with 10% FBS and 1% NEAA. For subculturing, cells are trypsinized and resuspended in EMEM supplemented with 10% FBS and 1% NEAA. Cells are seeded into new flasks at a density of 1 x 10⁴ cells per flask.

Seeding density 1 x 10⁴ cells / flask

Fluid renewal 2-3 times per week

Post-Thaw Recovery After thawing, cells are seeded into a 150 cm² flask containing 150 ml of EMEM supplemented with 10% FBS and 1% NEAA. Cells are allowed to recover for 24 hours before being passaged.

Freeze medium EMEM supplemented with 10% FBS and 1% NEAA (Freeze FBS) + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the vial containing the cells in a 37°C water bath.
 2. Centrifuge the cells at 300 x g for 3 minutes.
 3. Resuspend the cells in 15 ml of EMEM supplemented with 10% FBS and 1% NEAA.
 4. Seed the cells into a 150 cm² flask containing 150 ml of EMEM supplemented with 10% FBS and 1% NEAA.
 5. Allow the cells to recover for 24 hours before being passaged.
 6. Pass the cells into a new flask at a density of 1 x 10⁴ cells per flask.
 7. Repeat the process for subsequent passages.
 8. Store the cells in a liquid nitrogen storage tank.

