

Product sheet

HOS | 300449

General Information

| | |
|--------------------|---|
| Description | HOS (ATCC CCL-21) is a continuous cell line derived from a 13-year-old female patient with osteosarcoma. HOS cells are derived from the primary tumor and are characterized by their ability to form osteoid matrix in culture. HOS cells are a derivative of the HOS 143B cell line. |
| Organism | Human |
| Tissue | Osteosarcoma |
| Disease | Osteosarcoma |

Characteristics

| | |
|--------------------------|---|
| Age | 13 years |
| Gender | Female |
| Ethnicity | White |
| Morphology | Epithelial cells, adherent |
| Growth properties | Highly proliferative, anchorage dependent |

Identification

| | |
|-----------------------------|---------------------------------|
| Citation | HOS (ATCC CCL-21) Cytion 300449 |
| Biosafety level | 1 |
| NCBI_TaxID | 9606 |
| CellosaurusAccession | CVCL_0312 |

Enzymes

| | |
|-------------------|---------|
| Isoenzymes | G6PD, B |
|-------------------|---------|

HEK293T HOS | 300449

HEK293T

Culture Medium EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO₃, w: EBSS (Cytion Cytion 820100a)

Supplements Cytion Cytion 10% FBS 1% NEAA

Dissociation Reagent Cytion Cytion

Subculturing HEK293T cells are cultured in EMEM supplemented with 10% FBS and 1% NEAA. For subculturing, cells are trypsinized and resuspended in EMEM supplemented with 10% FBS and 1% NEAA. Cells are seeded into new flasks at a density of 1 x 10⁴ cells per flask.

Seeding density 1 x 10⁴ cells / flask

Fluid renewal 2-3 times per week

Post-Thaw Recovery After thawing, cells are cultured in EMEM supplemented with 10% FBS and 1% NEAA for 24 hours to allow recovery.

Freeze medium EMEM supplemented with 10% FBS and 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the vial in a 37°C water bath and transfer the cells to a 15 mL centrifuge tube.
 2. Centrifuge the cells at 300 x g for 3 minutes and remove the supernatant.
 3. Resuspend the cells in EMEM supplemented with 10% FBS and 1% NEAA.
 4. Seed the cells into a 10 mL flask at a density of 1 x 10⁴ cells per flask.
 5. Incubate the cells in a humidified CO₂ incubator at 37°C.
 6. Monitor the cells for attachment and growth.
 7. Once cells are attached, replace the medium with EMEM supplemented with 10% FBS and 1% NEAA.
 8. Continue to culture the cells in EMEM supplemented with 10% FBS and 1% NEAA.

