

HEP-2 B-LCL-HROC24 | 300861

HEP-2 B-LCL-HROC24

Description	B-LCL-HROC24 is a cell line derived from a patient with B-cell lymphoma. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma. The cell line is characterized by the presence of EBV (Epstein-Barr virus) and is highly tumorigenic. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma. B-LCL-HROC24 is a cell line derived from a patient with B-cell lymphoma. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma. The cell line is characterized by the presence of EBV (Epstein-Barr virus) and is highly tumorigenic. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma. B-LCL-HROC24 is a cell line derived from a patient with B-cell lymphoma. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma. The cell line is characterized by the presence of EBV (Epstein-Barr virus) and is highly tumorigenic. It is a B-cell derived cell line that is highly tumorigenic and is used for research in B-cell lymphoma.
Organism	Human
Tissue	B-cell lymphoma
Disease	B-cell lymphoma
Synonyms	B-LCL CO24, Bc HROC24

HEP-2 B-LCL-HROC24

Age	98 years
Gender	Male
Ethnicity	White
Morphology	Large, round, epithelial cells
Cell type	B-cell lymphoma
Growth properties	Highly tumorigenic

HEP-2 B-LCL-HROC24

Citation	B-LCL-HROC24 (HEP-2 B-LCL-HROC24 Cytion 300861)
Biosafety level	2
NCBI_TaxID	9606

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CellosaurusAccession CVCL_YD57

Cell Line HEp-2B-LCL-HROC24

Surface antigens CD19

Viruses EBV

Cell Line

Culture Medium RPMI 1640, w: 2.0 mM NaH_2PO_4 , w: 2.0 g/L NaHCO_3 (Cytion 820700a)

Supplements 10% FBS

Subculturing 1:5

Freeze medium 10% FBS + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a water bath at 37°C. Transfer the cells to a centrifuge tube and centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium.
2. Seed cells into a T25 flask at a density of 1×10^5 cells per flask. Incubate at 37°C with 5% CO_2 .
3. Monitor cell growth and confluency. Once cells reach 70-80% confluency, passage the cells.
4. Harvest cells by trypsinization and resuspend in 15 mL of complete medium. Seed into a T75 flask at a density of 1×10^5 cells per flask.
5. Monitor cell growth and confluency. Once cells reach 70-80% confluency, passage the cells.
6. Harvest cells by trypsinization and resuspend in 15 mL of complete medium. Seed into a T75 flask at a density of 1×10^5 cells per flask.
7. Monitor cell growth and confluency. Once cells reach 70-80% confluency, passage the cells.
8. Harvest cells by trypsinization and resuspend in 15 mL of complete medium. Seed into a T75 flask at a density of 1×10^5 cells per flask.

Incubation Atmosphere 37°C, 5% CO_2

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Flask Coating

Flask coating is not required for this product.

Freezing Procedure

For freezing, cells should be seeded at a density of 1 x 10⁶ cells per 100 cm² flask. After 24 hours, the medium should be replaced with fresh medium. The cells should be harvested after 48 hours and frozen in liquid nitrogen. The cells should be stored at -78°C.

Shipping Conditions

The cells should be shipped in a dry ice container. The shipping container should be labeled with the product name and the shipping conditions. The cells should be stored at -78°C.

Storage Conditions

The cells should be stored in a liquid nitrogen container. The storage container should be labeled with the product name and the storage conditions. The cells should be stored at -150 to -196 °C.

HLA-B-LCL-HROC24 / HLA

Sterility

The cells are sterile and free of mycoplasma contamination. The cells are tested for sterility using PCR and microscopy. The cells are also tested for mycoplasma contamination using PCR and microscopy. The cells are found to be sterile and free of mycoplasma contamination.

HLA

A*: '02:01:01, '30:02:01
B*: '18:01:01, '38:01:01
C*: 05:01:01, 12:03:01
DRB1*: 03:01:01, 13:01:01
DQA1*: '01:03:01, '05:01:01
DQB1*: '02:01:01, '06:03:01
DPB1*: '02:02:01, '03:01:01
E: '01:01:01, '01:03:02