



**HEK293T B82 | 305173**

**Culture Medium** DMEM, w: 4.5 g/L  $\beta$ -glucose, w: 4 mM L-glutamine, w: 3.7 g/L NaHCO<sub>3</sub>, w: 1.0 mM  $\beta$ -mercaptoethanol (Cytion 820300a)

**Supplements**  $\beta$ -glucose 10% FBS

**Dissociation Reagent**  $\beta$ -glucose

**Subculturing** HEK293T cells are cultured in DMEM supplemented with 10% FBS. For subculturing, cells are trypsinized and resuspended in DMEM supplemented with 10% FBS. Cells are seeded into new flasks at a density of 1 x 10<sup>6</sup> cells per flask.

**Fluid renewal** 2 x 3 days

**Freeze medium** DMEM supplemented with 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells rapidly in a 37°C water bath.
  2. Centrifuge cells at 300 x g for 3 minutes.
  3. Resuspend cells in DMEM supplemented with 10% FBS.
  4. Seed cells into new flasks at a density of 1 x 10<sup>6</sup> cells per flask.
  5. Incubate cells in a 37°C incubator with 5% CO<sub>2</sub>.
  6. Monitor cell growth and confluency.
  7. Perform subculturing when cells reach 70-80% confluency.
  8. Repeat the process for subsequent passages.

**Incubation Atmosphere** 37°C, 5% CO<sub>2</sub>

**Flask Coating**  $\beta$ -glucose

**Freezing Procedure** Cells are trypsinized and resuspended in DMEM supplemented with 10% FBS + 10% DMSO. Cells are seeded into new flasks at a density of 1 x 10<sup>6</sup> cells per flask.

