

Product sheet

NCI-H446 | 305049

General Information

Description	Cell line established in 1982 by D. Carney, A.F. Gazdar from a patient with squamous cell carcinoma of the lung. The cells were grown in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS), 5% human platelet-derived growth factor (hPDGF), 10% human epidermal growth factor (hEGF), 10% human fibroblast growth factor (hFGF), and 10% human insulin-like growth factor (hIGF).
Organism	Human
Tissue	Lung
Disease	Non-small cell lung carcinoma
Metastatic site	Adipose tissue
Synonyms	NCI-H446, H-446, NCI-446, NCIH446

Cell Characteristics

Age	61 years
Gender	Male
Ethnicity	White
Morphology	Epithelial
Growth properties	Adherent

References and Safety

Citation	NCI-H446 (ATCC CCL-1562) Cytion 305049
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1562

Additional Information

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Tumorigenic Yes, tumorigenic in vivo (subcutaneous injection of 10^6 cells into immunodeficient mice, resulting in tumor formation in SCID mice).

Characteristics

Culture Medium RPMI 1640, w: 2.0 mM L-glutamine, w: 2.0 g/L NaHCO3 (supplemented with Cytion 820700a)

Supplements 10% FBS, 2.5 µg/ml insulin, 10 µg/ml HEPES, 1.0 µg/ml selenium

Dissociation Reagent Trypsin

Subculturing Cells are typically subcultured every 15-20 days using PBS wash followed by trypsin treatment (3-5 min at 37°C).

Split ratio 1:3 to 1:4

Fluid renewal 2-3 times per week

Freeze medium RPMI 1640, 10% FBS, 10% DMSO (supplemented with FBS) + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells rapidly in a 37°C water bath, then transfer to a 15 mL centrifuge tube containing 10 mL of pre-warmed complete culture medium.
 2. Centrifuge at 300 x g for 3 minutes, discard supernatant, and resuspend cells in 10 mL of complete culture medium.
 3. Count cells using a hemacytometer, and seed into a 25 cm^2 flask with 10 mL of complete culture medium.
 4. Allow cells to attach for 24 hours, then replace medium with fresh complete culture medium (70% replacement).
 5. Once cells are established, passage into a new flask using trypsin treatment (15 min at 37°C).
 6. Seed cells at a density of 10^5 cells per flask in 10 mL of complete culture medium.
 7. Monitor cell growth and confluency, and passage when cells reach 70-80% confluency.
 8. Maintain cells in complete culture medium with 10% FBS and 10% DMSO.

Incubation Atmosphere 37°C, 5% CO2, humidified

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Flask Coating

Freezing Procedure

Shipping Conditions

Storage Conditions

/ / **HLA**

Sterility

STR

Amelogenin: x,x

CSF1PO: 13

D13S317: 8

D16S539: 12

D5S818: 11

D7S820: 10,11

TH01: 8,9,3

TPOX: 9,11

vWA: 18,19

D3S1358: 17

D21S11: 28

D18S51: 12,13

Penta E: 9,1

Penta D: 12,13

D8S1179: 13,15

FGA: 22

D1S1656: 14,16,3

D6S1043: 11

D2S1338: 18,2

D12S391: 17,18

D19S433: 13,14