

Product sheet

**U2OS-CRISPR-NUP96-SNAP | 300444**

**U2OS-CRISPR-NUP96-SNAP**

**Description** U-2 OS-CRISPR-NUP96-SNAP (U2OS-CRISPR-NUP96-SNAP) is a cell line derived from U-2 OS. It is a CRISPR-Cas9 edited cell line with a stable knock-out of NUP96. U-2 OS-CRISPR-NUP96-SNAP clone no. 33, SNAP NUP96 (U2OS-CRISPR-NUP96-SNAP clone no. 33) is a CRISPR-Cas9 edited cell line with a stable knock-out of NUP96. U-2 OS-CRISPR-NUP96-SNAP clone no. 33 is a CRISPR-Cas9 edited cell line with a stable knock-out of NUP96. NPC

**Organism** Human  
**Tissue** Cell Culture  
**Disease** Cancer

**Characteristics**

**Age** 15 days  
**Gender** Male  
**Ethnicity** Caucasian  
**Growth properties** Adherent

**References**

**Citation** U-2 OS-CRISPR-NUP96-SNAP (U2OS-CRISPR-NUP96-SNAP) Cytion 300444  
**Biosafety level** 1  
**NCBI\_TaxID** 9606  
**CellosaurusAccession** CVCL\_B7FL  
**Depositor** Cytion GmbH (EMBL)  
**GMO Status** GMO-S1: U2OS-CRISPR-NUP96-SNAP (U2OS-CRISPR-NUP96-SNAP, clone 33) NUP96-SNAP

**Additional information**

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**Protein expression** NUP96-SNAP (CRISPR-Cas9 mediated knock-in of NUP96, SNAP-tag)

**CRISPR**

**Culture Medium** McCoy's 5a, w: 3.0 g/L  $\beta$ -mercaptoethanol, w: 1000 mg/ml penicillin, w: 2.0 mM L-glutamine, w: 2.2 g/L NaHCO<sub>3</sub> (Cytion 820200a)

**Supplements** 10% FBS, 3.0 mg/ml insulin, 3.0 mg/ml transferrin, 2.0 mg/ml selenium, 2.2 mg/ml NaHCO<sub>3</sub>, 1% NEAA

**Dissociation Reagent** Trypsin

**Subculturing** Cells are cultured in McCoy's 5a medium supplemented with 10% FBS, 3.0 mg/ml insulin, 3.0 mg/ml transferrin, 2.0 mg/ml selenium, 2.2 mg/ml NaHCO<sub>3</sub>, and 1% NEAA. Cells are seeded into T25 flasks at a density of 1 x 10<sup>4</sup> cells per flask. Media is replaced every 3 days. Cells are passaged when they reach 70-80% confluency.

**Seeding density** 1 x 10<sup>4</sup> cells/flask

**Fluid renewal** 2-3 times per week

**Freeze medium** McCoy's 5a medium supplemented with 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells rapidly in a 37°C water bath. Transfer cells to a pre-warmed medium.
  2. Centrifuge cells at 300 x g for 3 minutes. Resuspend cells in 1 ml of fresh medium.
  3. Seed cells into T25 flasks at a density of 1 x 10<sup>4</sup> cells per flask.
  4. Allow cells to recover in fresh medium for 24 hours.
  5. After 24 hours, replace the medium with fresh McCoy's 5a medium supplemented with 10% FBS.
  6. Monitor cell growth and confluency. Media is replaced every 3 days.
  7. Cells are passaged when they reach 70-80% confluency.
  8. Cells are cultured in McCoy's 5a medium supplemented with 10% FBS, 3.0 mg/ml insulin, 3.0 mg/ml transferrin, 2.0 mg/ml selenium, 2.2 mg/ml NaHCO<sub>3</sub>, and 1% NEAA.

