

Product sheet

HEK293T FRTL-5 | 500407

Culture Medium Ham's F12, w: 1.0 mM β -mercaptoethanol, w: 1.0 mM β -mercaptoethanol, w: 1.1 g/L NaHCO₃ (Cytion 820600a)

Supplements 5% FBS, 10 ng/ml transferrin, 5 ng/ml selenium, 50 ng/ml insulin, 10 ng/ml hydrocortisone, 10 ng/ml prolactin, 10 ng/ml dexamethasone

Dissociation Reagent Trypsin

Doubling time 30-34 hours

Subculturing Seed cells into 96-well plates at 1000 cells/well in 100 µl of medium. For T25 flasks, seed 1-3 x 10⁶ cells in 100 ml of medium. For T75 flasks, seed 3-5 x 10⁶ cells in 300 ml of medium.

Freeze medium DMEM (10% FBS) + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells in a 37°C water bath. Transfer cells to a 15 ml centrifuge tube and add 10 ml of pre-warmed medium.
 2. Centrifuge cells at 300 x g for 3 minutes. Remove the supernatant and resuspend the cell pellet in 10 ml of pre-warmed medium.
 3. Seed cells into a T25 flask at 100,000 cells per flask. Incubate at 37°C in 5% CO₂.
 4. Once cells reach 70% confluency, passage them into a T75 flask.
 5. Seed cells into a T75 flask at 1.5 x 10⁶ cells per flask. Incubate at 37°C in 5% CO₂.
 6. Once cells reach 70% confluency, passage them into a T175 flask.
 7. Seed cells into a T175 flask at 1.5 x 10⁷ cells per flask. Incubate at 37°C in 5% CO₂.
 8. Once cells reach 70% confluency, passage them into a T250 flask.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating Poly-D-Lysine

