



**HEK293T L5178-R | 400258**

**Viruses**

Adenovirus MAP (Ad5, Ad6, Ad15, Ad35, Ad48, Ad55, Ad57, Ad59, Ad69, Ad72, Ad79, Ad119, Ad147, Ad161, Ad162, Ad163, Ad164, Ad165, Ad166, Ad167, Ad168, Ad169, Ad170, Ad171, Ad172, Ad173, Ad174, Ad175, Ad176, Ad177, Ad178, Ad179, Ad180, Ad181, Ad182, Ad183, Ad184, Ad185, Ad186, Ad187, Ad188, Ad189, Ad190, Ad191, Ad192, Ad193, Ad194, Ad195, Ad196, Ad197, Ad198, Ad199, Ad200), Adenovirus K, Adenovirus Reo 3, PVM, LCM, M.pulmonis, MVM, GD VII, Herpesvirus H-1, Herpesvirus 8

**Cell Line**

**Culture Medium**

RPMI 1640, w: 2.0 mM L-glutamine, w: 2.0 g/L NaHCO3 (Cytion 820700a)

**Supplements**

10% FBS, 1 mM NEAA

**Subculturing**

5-10% FBS, 5-10% NEAA, 5 x 10<sup>6</sup> cells

**Seeding density**

1 x 10<sup>6</sup> cells/cm<sup>2</sup>

**Fluid renewal**

3 days

**Post-Thaw Recovery**

2-4 days

**Freeze medium**

DMEM, 10% FBS + 10% DMSO

**Thawing and Culturing Cells**

1. Thaw the vial quickly in a 37°C water bath, and transfer the cells to a pre-warmed tube containing 10 ml of complete medium.
2. Centrifuge the cells at 300 x g for 3 minutes, remove the supernatant, and resuspend the cells in 10 ml of complete medium.
3. Seed the cells into a T25 flask containing 10 ml of complete medium.
4. Incubate the cells in a humidified CO<sub>2</sub> incubator at 37°C.
5. Once the cells have reached confluence, passage them into a new T25 flask.
6. Repeat the passage process every 2-3 days.
7. When the cells reach confluence, they can be used for experiments.
8. If you need to freeze the cells, follow the freezing protocol.

