

Product sheet

**HROG10 | 300935**

**General information**

<b>Description</b>	Cell line derived from a patient with glioblastoma (PD Dr. Michael Linnebacher)
<b>Organism</b>	Human
<b>Tissue</b>	Brain, Glioma
<b>Disease</b>	Glioblastoma (WHO grade IV)

**Characteristics**

<b>Age</b>	74 years
<b>Gender</b>	Male
<b>Ethnicity</b>	German
<b>Morphology</b>	Epithelial cells, adherent, growing in monolayers
<b>Growth properties</b>	Highly proliferative

**Identification**

<b>Citation</b>	HROG10 (ATCC CCL-222) Cytion 300935
<b>Biosafety level</b>	1
<b>NCBI_TaxID</b>	9606
<b>CellSaurusAccession</b>	CVCL_4U43

**Antigen expression and mutational profile**

<b>Antigen expression</b>	HLA-A02 +, HLA-B*08:01 + HLA-E +, HLA-G -, MIC A +, MIC-B -, ICAM-1 +, GFAP +, CD133 +, CD133 +, S-100+, GBM +, BT
<b>Mutational profile</b>	TP53 wt, PTENwt, 4q12 (PDGFRA) loss

### HEK293T | HROG10 | 300935

#### HEK293T

<b>Culture Medium</b>	DMEM:Ham's F12 (1:1), w: 3.1 g/L $\beta$ -mercaptoethanol, w: 2.5 mM L-glutamine, w: 15 mM HEPES, w: 0.5 mM $\text{CaCl}_2 \cdot \text{H}_2\text{O}$ , w: 1.2 g/L $\text{NaHCO}_3$ (820400a)
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<b>Supplements</b>	$\beta$ -mercaptoethanol 10% FBS
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<b>Dissociation Reagent</b>	Trypsin
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<b>Subculturing</b>	<p>HEK293T cells are cultured in DMEM:Ham's F12 (1:1) supplemented with 10% FBS and <math>\beta</math>-mercaptoethanol. For subculturing, cells are trypsinized and resuspended in DMEM:Ham's F12 (1:1) supplemented with 10% FBS and <math>\beta</math>-mercaptoethanol. Cells are seeded into T25 or 35 cm<sup>2</sup> flasks.</p>
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<b>Seeding density</b>	$1 \times 10^4$ cells/cm <sup>2</sup>
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<b>Fluid renewal</b>	3-5 days
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<b>Freeze medium</b>	DMEM:Ham's F12 (1:1) supplemented with 50% FBS + 40% DMSO, CM-1 (Cytion 800100), $\beta$ -mercaptoethanol
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- Thawing and Culturing Cells**
1. Thaw the cells in a 37°C water bath and transfer them to a 15 ml centrifuge tube. Add 10 ml of DMEM:Ham's F12 (1:1) supplemented with 10% FBS and  $\beta$ -mercaptoethanol.
  2. Centrifuge the cells at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM:Ham's F12 (1:1) supplemented with 10% FBS and  $\beta$ -mercaptoethanol.
  3. Seed the cells into a T25 flask containing 37 ml of DMEM:Ham's F12 (1:1) supplemented with 10% FBS and  $\beta$ -mercaptoethanol.
  4. Seed the cells at a density of  $1 \times 10^4$  cells/cm<sup>2</sup>. Once the cells reach 70% confluency, perform a split.
  5. Seed the cells into a T25 flask containing 37 ml of DMEM:Ham's F12 (1:1) supplemented with 10% FBS and  $\beta$ -mercaptoethanol.
  6. Seed the cells at a density of  $1 \times 10^4$  cells/cm<sup>2</sup>.
  7. Seed the cells into a T25 flask containing 37 ml of DMEM:Ham's F12 (1:1) supplemented with 10% FBS and  $\beta$ -mercaptoethanol.
  8. Seed the cells at a density of  $1 \times 10^4$  cells/cm<sup>2</sup>.

<b>Incubation Atmosphere</b>	37°C, 5% $\text{CO}_2$ , humidified
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