

Product sheet

HEK293T | 305133

Culture Medium DMEM, w: 4.5 g/L D-glucose , w: 4 mM L- glutamine , w: 3.7 g/L NaHCO_3 , w: 1.0 mM $\text{beta-mercaptoethanol}$ (Cytion 820300a)

Supplements $\text{beta-mercaptoethanol}$ 10% FBS

Dissociation Reagent $\text{beta-mercaptoethanol}$

Doubling time 22 - 30 hours

Subculturing Cells are grown in 96-well plates, 384-well plates, T25, 75 cm² flasks, 300 cm² bioreactors, and 1000 cm² bioreactors. Cells are grown in DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.

Fluid renewal 2 - 3 times per week

Freeze medium $\text{beta-mercaptoethanol}$ free medium, supplemented with 10% FBS + 10% DMSO (Cytion 820300a)

Thawing and Culturing Cells

1. Thaw the cells in a 37°C water bath.
2. Dilute the cells into a 150 cm² flask with 150 ml of DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.
3. Incubate the cells at 37°C in 5% CO_2 .
4. Once cells reach 70% confluency, passage the cells.
5. Seed cells into a 15 cm² flask with 15 ml of DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.
6. Once cells reach 70% confluency, passage the cells into a 300 cm² bioreactor with 300 ml of DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.
7. Seed cells into a 10 cm² flask with 10 ml of DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.
8. Once cells reach 70% confluency, passage the cells into a 1000 cm² bioreactor with 1000 ml of DMEM supplemented with 10% FBS and 1.0 mM $\text{beta-mercaptoethanol}$.

Incubation Atmosphere 37°C, 5% CO_2 , humidified

Flask Coating $\text{beta-mercaptoethanol}$ free medium, supplemented with 10% FBS + 10% DMSO (Cytion 820300a)

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PATU8988T | 305133

Freezing Procedure -78°C

Shipping Conditions -78°C

Storage Conditions -150 to 196

HLA

Sterility