

Product sheet

6T-CEM | 305132

6T-CEM

Culture Medium MEM, w: 2.0 mM β -mercaptoethanol, w/o: penicillin, w/o: streptomycin, w: 1.0 mM nystatin, w: 2.2g/ml insulin-like growth factor 1 (IGF1), w: 2.2g/ml transferrin, w: 2.2g/ml selenium

Supplements 10% FBS

Subculturing 1:5

Fluid renewal 2-3 times per week

Freeze medium MEM, w: 2.0 mM β -mercaptoethanol, w/o: penicillin, w/o: streptomycin, w: 1.0 mM nystatin, w: 2.2g/ml insulin-like growth factor 1 (IGF1) + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a 37°C water bath, then transfer to a 37°C incubator with 5% CO₂.
2. Allow cells to recover in the incubator for 24-48 hours before passaging.
3. After 24-48 hours, check for cell attachment. If cells are not attached, gently tap the flask and check again after 24 hours.
4. Once cells are attached, replace the medium with fresh medium.
5. Pass cells into a new flask when they reach 70-80% confluency.
6. Use a pipette to remove the medium and wash the cells with PBS.
7. Add 1-2 ml of trypsin to the flask and incubate for 5-10 minutes at 37°C.
8. Add 10 ml of medium to stop the trypsin reaction and pipette up the cells into a 15 ml centrifuge tube.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating None

Freezing Procedure Harvest cells into a 15 ml centrifuge tube, centrifuge at 300 x g for 5 minutes, remove the supernatant, wash with PBS, and resuspend in 1 ml of freezing medium. Store at -80°C.

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Shipping Conditions

Store at -78°C

Storage Conditions

Store at -150 to 196°C

HLA

Sterility

PCR