

Product sheet

XXXX HNO258 | 300146

XXXX XXXX

Description XXXX XXXX
XXXX HNO258 XXXX XXXXXXXXXXXX XX XXX XXXX XXXXXXXXXXXX, XXXX XX-XXXX XX XXXXXXXXXXXX XX XXX XXXX XXXX XXXXXXXXXXXX (HNSCC). XX XXXX p13,9q31-qter, 11q13, 15p X-15q. XXXXXXX, XXXX XXXX XXXXXXX XXXXXXX XXXXXXXXXXXX XXXXXXXXXXXX 4p X-18q12-qter. XXXXXXXXXXX XXXX XXXXXXX XXXXXXXXXXX XX 11q13, XXXXXXX X-HNO258, XXXXXXX XXXXXXX XXXXXXX XXX XXXX XXX XXXXXXXXXXXX XXXX CCND1 (XXXXXX D1) X-C

Organism XXXX

Tissue XXXX XXXX

Disease XXXXXXXXXXX XX XXX XXXX XXXX XXXXXXX (HNSCC)

XXXXXXXXXXXX

Age 62 XXXX

Gender XXXX

Ethnicity XXXXXXX

Morphology XXXX XXXXXXX

Growth properties XXXX XXX, XXXXXXX

XXXXXXXXX XXXXXXXXXXXXXXX

Citation HNO258 (XXXX XXXXXXX Cytion 300146)

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_D221

XXXXXXXXX XXXX-XXXXXXXXXXXX

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Product sheet

HEK293T HNO258 | 300146

Culture Medium DMEM, w: 4.5 g/L D-glucose, w: 4 mM L-glutamine, w: 3.7 g/L NaHCO₃, w: 1.0 mM β-mercaptoethanol (Cytion 820300a)

Supplements 10% FBS

Dissociation Reagent Trypsin

Subculturing Seed cells into fresh medium containing 10% FBS. For passage 2-3, use 1:3 split ratio. For passage 4-5, use 1:5 split ratio. For passage 6-7, use 1:10 split ratio. For passage 8-9, use 1:20 split ratio. For passage 10, use 1:50 split ratio.

Fluid renewal 2-3 times per week

Freeze medium DMEM + 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells in a 37°C water bath.
 2. Centrifuge cells at 300 x g for 3 minutes.
 3. Wash cells with PBS.
 4. Resuspend cells in fresh medium containing 10% FBS.
 5. Seed cells into a 15 cm² flask.
 6. Incubate cells at 37°C, 5% CO₂.
 7. Monitor cell growth and passage when cells reach 70-80% confluency.
 8. Perform subculturing as described in the Subculturing section.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating Not required

