

Product sheet

RT-112-D21 | 300325

Culture Medium RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a)

Supplements 10% FBS

Dissociation Reagent

Subculturing Cells are harvested by trypsinization with Trypsin-EDTA (Cytion 820700a) and centrifuged at 300 x g for 5 min. Cells are resuspended in DMEM supplemented with 10% FBS and seeded into new flasks.

Split ratio 1:4 to 1:8

Fluid renewal 2 to 3 times per week

Freeze medium DMEM supplemented with 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the vial quickly in a 37°C water bath and transfer the cells to a centrifuge tube.
 2. Centrifuge the cells at 300 x g for 5 min and resuspend in 1 mL of DMEM supplemented with 10% FBS.
 3. Seed the cells into a 25 cm² flask containing 10 mL of DMEM supplemented with 10% FBS.
 4. Incubate the cells at 37°C in 5% CO₂ until they reach 70-80% confluency.
 5. Harvest the cells by trypsinization with Trypsin-EDTA (Cytion 820700a) and centrifuge at 300 x g for 5 min.
 6. Resuspend the cells in 1 mL of DMEM supplemented with 10% FBS.
 7. Seed the cells into a 25 cm² flask containing 10 mL of DMEM supplemented with 10% FBS.
 8. Incubate the cells at 37°C in 5% CO₂ until they reach 70-80% confluency.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating

