

ATCC NCTC 1469 | 400300

Thawing and Culturing Cells

1. Thaw the vial rapidly in a water bath at 37°C. Do not allow the vial to warm to room temperature. Immediately transfer the cells to a pre-warmed T25 flask containing 10 mL of complete medium.
2. Allow the cells to settle for 10 minutes. Then, add 10 mL of complete medium to the flask. Incubate at 37°C in 5% CO₂.
3. After 24 hours, check the cells. If they are not attached, add another 10 mL of complete medium. If they are attached, wait 24 hours before adding more medium.
4. Once the cells are attached, they can be passaged. Remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution. Incubate at 37°C for 5 minutes.
5. Add 10 mL of complete medium to stop the trypsin. Pipette the cells into a 15 mL conical tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant.
6. Resuspend the cells in 1 mL of complete medium. Count the cells using a hemacytometer. Seed the cells into a new flask at a density of 1 x 10⁶ cells per flask.
7. Incubate the cells at 37°C in 5% CO₂. Change the medium every 2-3 days.
8. Once the cells are in the log phase, they can be used for experiments.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating None

Freezing Procedure Harvest cells at 70-80% confluency. Wash with PBS. Add 2 mL of freezing medium. Centrifuge at 300 x g for 5 minutes. Resuspend in 1 mL of freezing medium. Store at -80°C.

Shipping Conditions Ship at 4°C. Do not freeze. Do not expose to light.

Storage Conditions Store at -150°C. 196 hours. Store in liquid nitrogen.

ATCC 1469 / ATCC 400300 / HLA

Sterility The cells are free of mycoplasmas and other contaminants. PCR confirmed. No mycoplasma detected.

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██████ STR

- M_18-3: 16
- M_4-2: 20,3, 21,3
- M_6-7: 12,13
- M_3-2: 14,15
- M_19-2: 11,12
- M_7-1: 26,27
- M_1-1: 10
- M_8-1: 14,16
- M_2-1: 9
- M_15-3: 25 ██████
- M_6-4: 18
- M_11-2: 16,18
- M_1-2: 16
- M_17-2: 15, 16, 17
- M_12-1: 16
- M_5-5: 15
- M_X-1: 26
- M_13-1: 17,18
- Human D4/D8: -