

HuH7 | 300156

Thawing and Culturing Cells

1. Thaw the vial rapidly in a water bath at 37°C. Do not allow the cells to warm to room temperature. Transfer the cells to a pre-warmed T25 flask containing 10 ml of complete medium.
2. Allow the cells to settle for 15 minutes. Add 10 ml of complete medium to the flask. Incubate the cells at 37°C in 5% CO₂.
3. After 24 hours, check the cells under a microscope. If the cells are not attached, add another 10 ml of complete medium. If the cells are attached, wait 24 hours before adding more medium.
4. When the cells are attached, they will form a monolayer. Harvest the cells when they reach 70-80% confluency.
5. Seed the cells into a 15 cm² flask with 8 ml of complete medium.
6. Harvest the cells when they reach 300 x 10³ cells. Seed the cells into a 300 x g flask with 3 ml of complete medium.
7. Harvest the cells when they reach 10⁷ cells. Seed the cells into a 10 cm² flask with 10 ml of complete medium.
8. Harvest the cells when they reach 10⁸ cells. Seed the cells into a 10 cm² flask with 10 ml of complete medium.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating None

Freezing Procedure Harvest cells at 70-80% confluency. Wash cells with PBS. Add 1 ml of freezing medium. Resuspend cells in 1 ml of freezing medium. Freeze cells in a cryovial in a dry ice/ethanol slush. Store cells at -80°C.

Shipping Conditions Store cells at -80°C. Ship cells in a dry ice/ethanol slush.

Storage Conditions Store cells at -150°C for 196 days. Thaw cells in a water bath at 37°C.

Genotype / HLA

Sterility Cells are tested for mycoplasma contamination using PCR. Cells are also tested for endotoxin contamination. Cells are sterile.

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A*: 11:01:01

B*: '54:01:01

C*: 01:02:01

DRB1*: 08:03:02

DQA1*: 01:03:01

DQB1*: 06:01:01

DPB1*: 02:01:02