

Product sheet

HEK293T | 305080

Culture Medium RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a)

Supplements β -mercaptoethanol 10% FBS

Dissociation Reagent Trypsin

Subculturing Cells are cultured in DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin. For differentiation, cells are seeded in DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin. After 24 hours, the medium is replaced with DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin.

Fluid renewal 2 x 3 days

Freeze medium DMEM supplemented with 10% FBS, 10% DMSO, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin.

- Thawing and Culturing Cells**
1. Thaw cells in a 37°C water bath and transfer to a 15 ml centrifuge tube.
 2. Add 10 ml of DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin.
 3. Centrifuge at 300 x g for 5 minutes.
 4. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin.
 5. Seed cells into a 25 cm² flask.
 6. Incubate cells in a 37°C incubator with 5% CO₂.
 7. Monitor cell growth and passage cells when they reach 70-80% confluency.
 8. For differentiation, replace the medium with DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating Poly-D-Lysine

